# Closing the global $N_2O$ budget: nitrous oxide emissions through the agricultural nitrogen cycle

OECD/IPCC/IEA phase II development of IPCC guidelines for national greenhouse gas inventory methodology

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Key words: animal waste, fertilizer, greenhouse gas, inventory, nitrous oxide

#### **Abstract**

In 1995 a working group was assembled at the request of OECD/IPCC/IEA to revise the methodology for N<sub>2</sub>O from agriculture for the National Greenhouse Gas Inventories Methodology. The basics of the methodology developed to calculate annual country level nitrous oxide (N2O) emissions from agricultural soils is presented herein. Three sources of N<sub>2</sub>O are distinguished in the new methodology: (i) direct emissions from agricultural soils, (ii) emissions from animal production, and (iii) N<sub>2</sub>O emissions indirectly induced by agricultural activities. The methodology is a simple approach which requires only input data that are available from FAO databases. The methodology attempts to relate N<sub>2</sub>O emissions to the agricultural nitrogen (N) cycle and to systems into which N is transported once it leaves agricultural systems. These estimates are made with the realization that increased utilization of crop nutrients, including N, will be required to meet rapidly growing needs for food and fiber production in our immediate future. Anthropogenic N input into agricultural systems include N from synthetic fertilizer, animal wastes, increased biological N-fixation, cultivation of mineral and organic soils through enhanced organic matter mineralization, and mineralization of crop residue returned to the field. Nitrous oxide may be emitted directly to the atmosphere in agricultural fields, animal confinements or pastoral systems or be transported from agricultural systems into ground and surface waters through surface runoff. Nitrate leaching and runoff and food consumption by humans and introduction into sewage systems transport the N ultimately into surface water (rivers and oceans) where additional  $N_2O$  is produced. Ammonia and oxides of N (NO<sub>x</sub>) are also emitted from agricultural systems and may be transported off-site and serve to fertilize other systems which leads to enhanced production of N<sub>2</sub>O. Eventually, all N that moves through the soil system will be either terminally sequestered in buried sediments or denitrified in aquatic systems. We estimated global N<sub>2</sub>O-N emissions for the year 1989, using midpoint emission factors from our methodology and the FAO data for 1989. Direct emissions from agricultural soils totaled 2.1 Tg N, direct emissions from animal production totaled 2.1 Tg N and indirect emissions resulting from agricultural N input into the atmosphere and aquatic systems totaled 2.1 Tg N<sub>2</sub>O-N for an annual total of 6.3 Tg N<sub>2</sub>O-N. The N<sub>2</sub>O input to the atmosphere from agricultural production as a whole has apparently been previously underestimated. These new estimates suggest that the missing  $N_2O$  sources discussed in earlier IPCC reports is likely a biogenic (agricultural) one.

#### Introduction

The United Nations Framework Convention on Climate Change requires that all parties periodically update and publish national inventories of anthropogenic emissions by sources and removals by sinks of all greenhouse gases not controlled by the Montreal Protocol, using comparable methodologies. In response to this mandate the Intergovernmental Panel on Climate Change (IPCC), through the Office of Economic Cooperation and Development (OECD) and International Energy Agency (IEA) has been coordinating the development and updating of national inventory methodologies for various greenhouse gases. The first phase of methodology development was published in the 1995 IPCC Guidelines for National Greenhouse Gas Inventories (IPCC, 1995b). In phase II a working group of 32 persons from 18 countries was assembled at the request of OECD/IPCC/IEA to revise the National Greenhouse Gas Inventories Methodology for N<sub>2</sub>O from Agricultural Soils (IPCC, 1997). This paper presents the framework behind the phase II methodology for calculating annual, country level N<sub>2</sub>O emissions from agricultural soils (IPCC, 1997).

During the past decade attempts to define budgets for global atmospheric N2O suggested that the strength of known N2O sources is underestimated or that unidentified sinks exists (IPCC, 1990, 1992; Duxbury and Mosier, 1993; Robertson, 1993). In these budgeting efforts anthropogenic N2O emissions due to agricultural activities were considered to be relatively small (Table 1). These assessments were based upon a few reviews and interpretations that needed further examination (IPCC, 1992; Mosier, 1994; Mosier et al., 1996). Questions to these interpretations were beginning to be raised when the 1995 IPCC Guidelines for National Inventory Methodology for N<sub>2</sub>O in Agriculture (IPCC, 1995b) was being developed (Duxbury and Mosier, 1993; Mosier, 1994; Mosier and Bouwman, 1993). Before that time N<sub>2</sub>O emissions from agricultural systems were only considered from the aspect of direct N<sub>2</sub>O emissions from agricultural fields (OECD/OCDE, 1991) that had been fertilized with synthetic nitrogen (N) fertilizer. The estimates used tended to underestimate total agricultural emissions (Mosier, 1994; Bouwman, 1996). For example, animal production systems have the potential for significant N<sub>2</sub>O production (DeKlein and Logtenstijn, 1994) and needed consideration as did the remainder of the agricultural N cycle.

N applied to agricultural soils may be lost from the fields through surface erosion or leaching (Duxbury and Mosier, 1993). This leached N continues recycling in the soil–water–air system and eventually is denitrified and converted to  $N_2O$  and  $N_2$  and released back to the atmosphere (Figure 1; Nevison et al., 1996; Oonk and Kroeze, 1998), or buried in sediments. All of these pathways and factors needed to be included in the anthropogenic agricultural soil  $N_2O$  source.

The IPCC, 1995 Guidelines (IPCC, 1995b) included N2O emissions occurring directly from agricultural fields. The N sources in this calculation were expanded to include synthetic fertilizers, organic N from animal excreta and crop residue and the amount of biological N fixation. This basic formula equating direct N2O emissions from agricultural soils to the N input multiplied by a conversion factor of  $1.25 \pm 1.0\%$ was used in the Cole et al. (1996) Climate Change 1995 assessment of mitigation options for N<sub>2</sub>O emissions from agriculture. The derivation of this factor is further described in Mosier et al. (1996). Values from these estimates were included in the Climate Change 1994 (IPCC, 1994) report. Cole et al. (1996) included an additional factor of 0.75% of N applications to provide some accounting for indirect N2O emissions that eventually evolved back to the atmosphere from N leaching or runoff from agricultural fields as well as NO<sub>x</sub> and NH<sub>3</sub> volatilization (Cole et al., 1996; Mosier et al., 1996) (Table 1).

The IPCC 1995 Guidelines still lacked mechanisms for estimating N-fixation and crop residue input and a quantifiable method for calculating  $N_2O$  productions following N leaching and runoff. Additionally, animal production systems were not included in the agricultural anthropogenic  $N_2O$  production guidelines. As a start in overcoming these deficiencies in National Inventory estimates, we developed a Phase II method for estimating country scale anthropogenic  $N_2O$  emissions from agricultural soils. This paper describes the essence of the methodology developed and presents calculations which suggest that an underestimation of total anthropogenic  $N_2O$  emissions from agricultural systems is responsible for the previous imbalanced global  $N_2O$  budgets.

# Phase II Development of IPCC Guidelines

In 1993, the Phase I OECD/IPCC/IEA Guidelines workgroup (IPCC, 1995b) suggested that 'improving methodology for estimating N<sub>2</sub>O emissions may evolve in a series of steps, beginning with the N

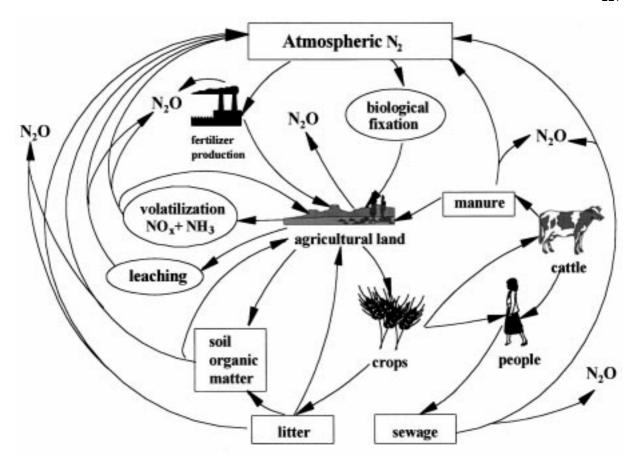


Figure 1. Depiction of the nitrogen cycle of agricultural soils and its relationship to N<sub>2</sub>O production. (Adapted from Nevison et al., 1996; Oonk & Kroeze (1998) by permission of John Wiley & Sons, Inc.).

source based equations and ending with development of process based models which are used to develop regional and larger scale emission models'. Using this recommendation and others noted in IPCC (1995b) the Phase II IPCC/OECD/IEA work group on Agricultural Soils initiated an effort to provide a more comprehensive  $N_2O$  emission calculation methodology.

As a first step, the Phase II workgroup evaluated the IPCC Guidelines (1995b) and recommended that the emission factor relating  $N_2O$  emissions directly from the soil to fertilizer N application should be that used by Cole et al. (1996) and Mosier et al. (1997). This value,  $1.25 \pm 1.0\%~N_2O$ –N of fertilizer N applied (Bouwman, 1994, 1996; Cole et al., 1996; Mosier et al., 1996, 1997) was derived from field information. The range of these values cover more than 90% of the published field data summarized by Bouwman (1994, 1996). Most of the information in Bouwman's summary was derived from field studies conducted in temperate regions of the world since few annual flux

measurements have been made in tropical agricultural systems.

The second step was to develop a more comprehensive methodology which attempted to take into account the major inputs of N into agriculture, include animal sources of  $N_2O$  and start to account for the indirect production of  $N_2O$  from N released from agriculture. This methodology does not account for the impact of climate, soils, and cropping system on  $N_2O$  production, consumption and emissions, because insufficient information is available with which to define appropriate emission coefficients for each variable. We defined some general principles that were used in methodology development:

- Use input data that are generally available worldwide. The data sources that best meet this requirement appear to be the agricultural yearbooks compiled by the United Nations Food and Agricultural Organization (FAO).
- 2) The general conceptual approach was adopted, in which fertilizer, animal waste, N derived from N<sub>2</sub>-

fixation, and crop residue N inputs to agricultural on an annual basis are tracked from their initial application to their return to the atmosphere via denitrification (Figure 1). All nitrogen flux estimates at various stages of the cycle should be consistent with the original N input and care must be taken to ensure that there is no double counting of N sources. The Phase II methodology is a first step towards such an integrated approach which assumes that all N input is reacted within one year and does not account for potential sequestration in the soil which may be released over decade to century time periods.

This new approach to estimating  $N_2O$  emissions from agricultural systems includes: (1) direct emissions of  $N_2O$  from agricultural fields ( $N_2ODIRECT$ ); (2) direct emissions of  $N_2O$  in animal production systems ( $N_2OANIMALS$ ) and (3) some of the indirect emission of  $N_2O$  that are derived from N that originated from agricultural systems ( $N_2OINDIRECT$ ). The general equation that is the basis for the calculation development described herein follows:

Total  $N_2O-N$  emissions from a country (kg  $N_2O-N$  y<sup>-1</sup>) are:  $N_2O=N_2ODIRECT+N_2OANIMALS+$   $N_2OINDIRECT$ 

## 2. Direct N<sub>2</sub>O emissions from agricultural soils

The IPCC Guidelines for National Greenhouse Gas Inventories only cover anthropogenic sources (i.e. the result of human activities). Anthropogenic sources of N<sub>2</sub>O can be biogenic (e.g. enhanced N<sub>2</sub>O production by bacteria in fertilized fields) or abiogenic (e.g. formation during burning processes). Several studies indicate that anthropogenic sources of N<sub>2</sub>O are largely biogenic, with agriculture as a major contributor (IPCC, 1995a; Bouwman et al., 1995; Mosier et al., 1996).

Biogenic production of  $N_2O$  in the soil results primarily from the nitrification and denitrification processes. Simply defined, nitrification is the aerobic microbial oxidation of ammonium to nitrate and denitrification is the anaerobic microbial reduction of nitrate to dinitrogen gas. Nitrous oxide is a gaseous intermediate in the reaction sequences of both processes which leaks from microbial cells into the soil atmosphere (Firestone and Davidson, 1989). Major

Table 1. Global  $N_2O$  budgets: IPCC (1992), IPCC (1994) and from the  $N_2O$  methodology presented in this paper for  $N_2O$  from cultivated soils (IPCC, 1997)

	IPCC, 1992	IPCC, 1995a	IPCC, 1997
Sources		$Tg N y^{-1}$	
Natural <sup>a</sup>			
ocean	1.4-2.6	3(1–5)	3.0(1-5)
tropical soils			
wet forest	2.2 - 3.7	3(2.2–3.7)	3.0(2.2-3.7)
dry savanas	0.5-2.0	1(0.5–2.0)	1.0(0.5-2.0)
temperate soils			
forests	0.5 - 2.0	1(0.1–2.0)	1.0(0.1–2.0)
grasslands	?	1(0.5–2.0)	1.0(0.5-2.0)
Subtotal	4.6-8.3	9(4.3–14.7)	9.0(4.3–14.7)
Anthropogenic			
agricultural soils	0.03 - 3.0	3.5(1.8–5.3)	$3.3^b(0.6-14.8)$
biomass burning	0.2 - 2.1	0.5(0.2-1.0)	0.5 (0.2–1.0)
industrial sources	0.8 - 1.8	1.3(0.7–1.8)	1.3 (0.7–1.8)
cattle and feedlots	?	0.4(0.2–0.5)	2.1 (0.6–3.1)
Subtotal	1.0-6.9	5.7(3.7–7.7)	7.2 (2.1–19.7)
Total sources	5.6-15.2	14.7(8-22.4)	16.2(6.4–34.4)
Sinks			
Atmospheric increase	3-4.5	.9(3.1–4.7)	3.9(3.1-4.7)
Soils	?	?	?
Stratospheric sink	7–13	12.3(9-16)	12.3(9–16)

<sup>&</sup>lt;sup>a</sup>For IPCC,1997 estimates of Natural N<sub>2</sub>O sources we use the 'likely' values from IPCC, 1995a. The values in parenthese in this column represent the range of estimates for each category.

regulators of these processes are carbon and nitrogen substrate availability, temperature, pH and soil moisture content.

In most agricultural soils biogenic formation of  $N_2O$  is enhanced by an increase in available mineral N which, in turn increases nitrification and denitrification rates. Addition of fertilizer N, therefore, directly results in extra  $N_2O$  formation (Figure 1). Most studies on  $N_2O$  emissions from agricultural soils investigate the difference in  $N_2O$  production between fertilized and unfertilized fields. Emissions from unfertilized fields are considered background emissions. However, actual background emissions from agricultural soils may be higher than historic natural emissions as a result of enhanced mineralization of soil organic matter

column represent the range of estimates for each category.  $^b$ The 3.3 shown here is 0.9 lower than the total in Table 11, because we assume that part of the natural soil and ocean emissions estimates include part of the indirect  $N_2O$  that we calculate from emissions of  $NH_3$  and  $NO_x$  from fertilization of agricultural soils and from nitrate leaching and runoff from these soils. The cattle and feedlot category is the animal production category from our estimates listed in Table 7.

due to previous agricultural activities. This is particularly observed in organic soils in both cold and warm climates over the globe (Bouwman and Van der Hoek, 1991; Kroeze, 1994). Background emissions may also be lower than historic emissions due to depletion of soil organic matter (Groffman et al., 1993).

# Sources of N<sub>2</sub>O directly related to N input into agricultural soils

There are a variety of sources of N in agricultural systems that we term anthropogenic which include: (A) synthetic fertilizers, (B) animal manures (urine and feces), (C) N derived from enhanced biological N-fixation (BVF) through  $N_2$ -fixing crops (D) crop residue returned to the field after harvest and (E) human sewage sludge application. Although some part of the animal manure N, crop residue and sewage may have come from previous application of synthetic fertilizer, the reentry of this N back into the soil systems renders it again succeptable to microbial processes which produce  $N_2O$ .

# A. Synthetic fertilizers and B. Animal excreta N used as fertilizer

Although synthetic fertilizers and animal manures are important sources of  $N_2O$ , their soil input is required to provide the N needed to meet global food production demands. The amount of synthetic fertilizer N applied to agricultural fields world-wide is well documented in the FAO data base (FAO Annual Yearbooks; or world wide web:

http://ww.fao.org/waicent/Agricul.htm). Although the amount of N used as fertilizer from animal excreta is more uncertain, estimates can be made, based on animal population and agricultural practices (IPCC, 1995b; Table 5). To account for the loss of fertilizer from NH<sub>3</sub> volatilization and emission of nitric oxide (NO) through nitrification after fertilizer is applied to fields, an NH<sub>3</sub> volatilization and NO emission factor is needed. Even though climate, soil, fertilizer placement and type, and other factors influence NH<sub>3</sub> volatilization and  $NO_x$  emission a fixed, default emission factor of 0.1 (kg NH<sub>3</sub>-N + NO<sub>x</sub>-N emitted/kg N excreted) is used for synthetic fertilizers and 0.2 (kg NH<sub>3</sub>-N + NO<sub>x</sub>-N emitted/kg N applied) for animal waste fertilizer (Table 3) (0.2 is used for animal waste because of the potentially larger NH<sub>3</sub> volatilization). The amount of N from these sources available for conversion to N<sub>2</sub>O is therefore equal to 90% of the synthetic fertilizer N applied and 80% of the animal waste N applied (Schepers and Mosier, 1991).

#### C. Biological N fixation

Both the amount of N fixed by biological N fixation in agricultural systems and the  $N_2O$  conversion coefficient are uncertain. Biological nitrogen fixation (BNF) supplies globally some 90 to 140 Tg N yr $^{-1}$  to agricultural systems (Peoples et al., 1995). Although more verification on these figures is necessary, most indications are that BNF contributes more N for plant growth than the total amount of synthetic N fertilizers applied to crops each year (Danso, 1995). The Phase I IPCC Guidelines (IPCC, 1995b) mention about equal rates. On average, BNF supplies 50–60% of the N harvested in grain legumes, 55–60% of the N in nitrogen fixing trees and 70–80% of the N accumulated by pasture legumes (Danso, 1995). Cultivation of grain legumes, however, often results in net soil N depletion.

In the tropics and subtropics, the use of Azolla (a genus of aquatic ferns which contains an  $N_2$ -fixing cyanobacterium) is widespread. Azolla fixes 20–25 kg N ha<sup>-1</sup> (Kumarasinghe and Eskew, 1991) which is released upon death and decomposition. This N serves to fertilize an associated crop and eventually stimulate  $N_2O$  formation.

Galbally et al. (1992) and Bouwman and Sombroek (1990) indicate that legumes may contribute to N<sub>2</sub>O emission in a number of ways. Atmospheric N<sub>2</sub> fixed by legumes can be nitrified and denitrified in the same way as fertilizer N, thus providing a source of N2O. Additionally, symbiotically living Rhizobia in root nodules are able to denitrify and produce N2O (O'Hara and Daniel, 1985). Galbally et al. (1992) suggest an emission rate of 4 kg N ha<sup>-1</sup> y<sup>-1</sup> for improved pastures, and Duxbury et al. (1982) suggest that legumes can increase N2O emissions from pastures by a factor of 2 or 3. More recently Carran et al. (1995) found annual N<sub>2</sub>O emissions ranging from 0.5 to 5 kg N<sub>2</sub>O-N depending upon the relative fertility of the sampling location. In old and young ryegrass/clover pastures Muller (personal communication) observed N<sub>2</sub>O emissions of 0.7 and 0.3 kg N  $ha^{-1} yr^{-1}$ , respectively.

Because of the uncertainty in knowing the amount of  $N_2$  fixed during N-fixation (Peoples et al., 1995) and the lack of country data on N-fixing crops, it is difficult to assign a conversion factor to  $N_2O$  emission that is related to the amount of N fixed by a crop. Total N input (FBN) is estimated by assuming that total crop biomass is about twice the mass of edible crop

(FAO, 1990b), and a certain N content of N fixing crop (FRACNRBF, Table 3). This crop production is defined in FAO crop data bases as pulses and soybeans. The N-fixation contribution does not include N<sub>2</sub>O produced in legume pastures. This N<sub>2</sub>O production is at least partially accounted for emissions from pastures that are being grazed. Australia and New Zealand, for example, contain large areas of pasture land that includes legumes as part of the pastoral system. Little data are available for other parts of the globe (Mosier et al., 1997).

#### D. Crop residue and E. Sewage sludge application

There is only limited information concerning reutilization of N from crop residues and N from sewage sludge applied to agricultural lands. Although the amount of N that recycles into agricultural fields through these mechanisms may add 25–100 Tg of N yr $^{-1}$  of additional N into agricultural soils (mainly from crop residues) the amount converted to N<sub>2</sub>O is not known. To account for the N<sub>2</sub>O in the inventory budget at this time the emission factor for fertilizers is used as default and the amount of N reentering cropped fields through crop residues is calculated from the FAO data concerning crop production.

Nitrous oxide emissions associated with crop residue decomposition are calculated here by estimating the amount of N entering soils as crop residue (FCR). The amount of nitrogen entering the crop residue pool is calculated from crop production data. Since FAO data only represent the edible portion of the crop, these must be roughly doubled to estimate total crop biomass. We assume a nitrogen percentage (FRACNCRBF and FRACNCR0; Table 3) to convert from kg dry biomass yr<sup>-1</sup> to kg N yr<sup>-1</sup> in crops. Some countries may have sufficient information to define the N content of crop biomass more precisely. As a default we suggest distinguishing between N-fixing crops (pulses and soybeans) and non-N-fixing crops. Some of the crop residues is removed from the field as crop (approximately 45%), and some may be burned (approximately 25% of the remaining residue in developing countries), or fed to animals. The amount of N in crop residue actually returned to a field is uncertain, as is the amount of time required for the N to mineralize. We assume here that input and impact on N<sub>2</sub>O production occur annually. Neither the amount of root biomass remaining in the soil nor the amount of plant residue fed to animals is accounted for in this crop residue estimate.

Because no appropriate estimates of sewage sludge N used as fertilizer were found this N input is not discussed further.

Agricultural systems which may represent unusually high  $N_2O$  sources or sinks

### Glasshouse farming

N-fertilizer application to glasshouse-grown crops are typically high (Postma et al., 1994). The available data are limited in scope, but three sets of studies indicate that N<sub>2</sub>O emissions from glasshouse crops are similar to those from fields per unit of N input. Postma et al. (1994) quantified NH<sub>3</sub> and N<sub>2</sub>O emissions from glasshouse cultivation of lettuce on a sandbed and found that NH<sub>3</sub> emissions and N<sub>2</sub>O emitted directly or in drainage water totaled less than 1% of the N applied. Daum (personal communication) measured N<sub>2</sub>O emissions from soilless culture cucumbers and found that N-loss rates as N2O ranged between 0.4 and 0.9% of the N input into the culture system. Pollaris (1994) measured N2O emission in a glasshouse cultivation of tomato and lettuce and found, respectively, 0.7 and 1.4% of the applied N emitted as N<sub>2</sub>O. Overall, these data suggest that N2O emissions from glasshouse agriculture do not need to be included separately in N2O emission inventories and should be included only in the total fertilizer N consumed within each country. The importance of another factor, N2O emission during steam disinfection of glasshouse soils, is uncertain. Postma et al. (1994) found that  $2-25 \text{ kg N}_2\text{O}-\text{N ha}^{-1}$ were lost during 10-h following soil steaming. The extent of glasshouses to which this practice is applied is not known.

### Cultivation of high organic content soils

Large  $N_2O$  emissions occur as a result of drainage and cultivation of organic soils (Histosols) due to enhanced mineralization of old, N-rich organic matter (Guthrie and Duxbury, 1978; Koops et al., 1996, 1997; Nykanen et al., 1995; Martikainen et al., 1996; Velthof et al., 1996a). The rate of N-mineralization is determined by the N-quality of the Histosol, drainage, management practices and climatic conditions. The range for enhanced emissions of  $N_2O$  due to cultivation is estimated to be 2-15 kg  $N_2O-N$  ha<sup>-1</sup> yr<sup>-1</sup> of cultivated Histosol. Default emission values of 5 kg  $N_2O-N$  ha<sup>-1</sup> yr<sup>-1</sup> are used for boreal and temperate regions and 10 kg  $N_2O-N$  ha<sup>-1</sup> yr<sup>-1</sup> for tropical regions (Table 2).

Soil sink for  $N_2O$ 

Aerobic soils are typically sources for N2O, but small uptake rates have been observed in isolated instances in dry soils (Duxbury and Mosier, 1993) and in wet grass pastures (Ryden, 1981, 1983). In a seasonally burned 'cerrado' in Brazil, Nobre (1994) observed occasional small but inconsistent consumption rates and concluded that this sink was very small in these soils. Anaerobic soils have a large potential for reducing N2O to N2 (Erich et al., 1984), since the major product of denitrification in soils is usually N2 rather than N2O. However, no large, constant N2O uptake has been reported and flooded rice fields (Parashar, 1991), for example, generally show very small emissions, depending upon the time of cropping season (Minami and Fukushi, 1984). Apparently slow rates of dissolution and transport of atmospheric N2O in wet/or flooded soils prevents this process from being a significant regulator of atmospheric N<sub>2</sub>O. Until additional information is available to indicate that soil uptake, in aerobic or flooded soils, is important, soil uptake of atmospheric N2O will not be included in the N<sub>2</sub>O budget for agricultural systems.

Methodology for estimating direct  $N_2O$  emissions from agricultural fields

Rather than repeating background information that has been published several times, we refer the reader to general discussions of conversion of different N sources to N<sub>2</sub>O and background materials to Bouwman (1994, 1995, 1996); Mosier (1994); Mosier et al. (1996, 1997) and IPCC Guidelines (IPCC, 1995b). The following methodology for assessing direct N<sub>2</sub>O emissions from agricultural fields includes consideration of synthetic fertilizer, N from animal waste, enhanced N<sub>2</sub>O production due to biological N-fixation, N from crop residue mineralization and soil N mineralization due to cultivation of Histosols. In this estimate the total direct annual N<sub>2</sub>O emission is:

$$N_2$$
ODIRECT = [(FSN + FAW + FBN + FCR)  
\* EF1] + FOS \* EF2

where

FSN = NFERT \* (1-FRACGASF)

FAW = (NEX \* (1-(FRACFUEL + FRACGRAZ + FRACGASM))

FBN = 2 \* CROPBF \* FRACNCRBF

FCR = 2 \* [CROP0 \* FRACNCR0 + CROPBF \* FRACNCRBF] \* (1-FRACR) \* (1-FRACBURN)

and

CROPBF = seed yield of pulses + soybeans in country (kg dry biomass  $yr^{-1}$ )

CROP0 = production of all other crops in country (kg dry biomass yr<sup>-1</sup>)

EF1 = Emission Factor for direct soil emissions (kg  $N_2O-N/kg\ N$  input); Table 2

EF2 = Emission Factor for organic soil mineralization due to cultivation (kg  $N_2O-N$  ha<sup>-1</sup> yr<sup>-1</sup>); Table 2

FAW = animal waste N used as fertilizer in country (kg N yr<sup>-1</sup>)

FBN = N fixed by N-fixing crops in country (kg N  $yr^{-1}$ )

FCR = N in crop residues returned to soils in country (kg N yr<sup>-1</sup>)

FOS = area of cultivated organic soils within a country (ha of Histosols in FAOdata base)

FRACBURN = fraction of crop residue that is burned rather than left on field; Table 3

FRACGASF = fraction of synthetic fertilizer N applied to soils that volatilizes as  $NH_3$  and  $NO_x$  (kg  $NH_3$ –N and  $NO_x$ –N/kg of N input); Table 3

FRACFUEL = fraction of livestock N excretion contained in excrements burned for fuel (kg N/Kg N total excreted);

FRACGASM = fraction of livestock N excretion that volatilizes as NH<sub>3</sub> and NO<sub>x</sub> (kg NH<sub>3</sub>–N and NO<sub>x</sub>–N/kg of N excreted); Table 3

FRACGRAZ = fraction of livestock N excretion contained in excrements deposited during grazing (kg N/kg N totally excreted); Appendix 1

FRACNCRBF = fraction of N in N-fixing crop (kg N/kg of dry biomass); Table 3

FRACNCR0 = fraction of N in non-N-fixing crop (kg N/kg of dry biomass); Table 3

FRACR = fraction of crop residue that is removed from the field as crop (kg N/kg crop-N); Table 3

 $FSN = synthetic N applied in country (kg N yr^{-1})$ 

 $N_2 ODIRECT = direct \ N_2 O \ emissions \ from \ agricultural soils in country (kg N \ yr^{-1})$ 

NEX = amount of N excreted by the livestock within a country (kg N yr<sup>-1</sup>); Table 5

NFERT = synthetic fertilizer use in country (kg N  $yr^{-1}$ )

The input data needed for this methodology include synthetic fertilizer use (NFERT), manure-N used as fertilizer (FAW), edible crop production of N-fixing crops (CROPBF) and non-N-fixing crops (CROPO), and area of cultivated organic soils (Histosols) in the country. The data for synthetic fertilizer

Table 2. Summary of default emission factors for agricultural emissions of N<sub>2</sub>O

$$\begin{split} & EF1 = 0.0125~(0.0025-0.0225)~kg~N_2O-N/kg~N~input\\ & EF2 = 5~temperate~and~10~tropical~(2-15)~(kg~N~ha^{-1}~yr^{-1})\\ & EF3:~see~Table~6\\ & EF4 = 0.01~(0.002-0.02)~kg~N_2O-N/kg~NH_3-N~and~NO_X-N~emitted\\ & EF5 = 0.025~(0.002-0.12)~kg~N_2O-N/kg~N~leaching/runoff\\ & EF6 = 0.01~(0.002-0.12)~kg~N_2O-N/kg~sewage-N~produced \end{split}$$

use are available on a country basis in the FAO data base (e.g. FAO, 1990a) and the amount of N in animal waste applied to agricultural fields (FAW) is calculated from the number and type of animals within a country (FAO data base; IPCC, 1995b) and an in-country estimate of the percentage of N excreted by farm animals that is collected during confinement and reapplied to the field (Table 5 and Appendix 1). Both synthetic fertilizer and manure used as fertilizer need to be corrected for the amount of NH<sub>3</sub> volatilized and NO<sub>x</sub> emitted (10 and 20% of N applied or excreted, respectively) after the material is placed in or on the soil so that the same N atom is not counted again. The FAW data also need to be carefully evaluated for each country to be sure that animal waste used to fertilize crops and animal waste deposited on pastures while animals are grazing are not double counted. Crop production data for pulses and soybeans and non-N-fixing crops are listed in the FAO crop data base (FAO, 1990b).

#### 3. Direct N<sub>2</sub>O emissions in animal production

#### N<sub>2</sub>O Sources within animal production systems

Earlier IPCC estimates (IPCC, 1995a) of  $N_2O$  emission from agriculture and other sources (IPCC, 1990, 1992) did not include  $N_2O$  emission from animal production. Recent studies (e.g. Bouwman, 1995; Jarvis and Pain, 1994; Flessa et al., 1996; Mosier et al., 1996, 97) indicate that emissions from animal wastes can be significant. There are three potential sources in animal production, i.e. (A) animals themselves, (B) wastes from confined animals and (C) dung and urine deposited on the soil by grazing animals. Emissions induced by use of manure N as fertilizer (with the exception of grazing animals) are considered direct  $N_2O$  emissions from agricultural fields and are included in the previous section of this paper.

#### A. N<sub>2</sub>O from animals

Animals themselves may be very small sources of N<sub>2</sub>O. Animal fodders contain 10 to 40 g of N/ kg dry matter. The greater part of this N is organically bound, but as total N content increases so does the nitrate (NO<sub>3</sub><sup>-</sup>) content, generally. Nitrate contents in fodders generally range from 1-10 g N/kg dry matter (Spoelstra, 1985). Upon passage through the digestive track of the animal, nitrate is reduced via dissimilatory nitrate reduction to NH<sub>3</sub>/NH<sub>4</sub><sup>+</sup>. The nitrate reduction reaction may release small amounts of N2O in the gut (Kaspar and Tiedje, 1981), which may escape to the atmosphere during rumination. Though this possible route of N2O formation has been known for over 10 years, quantitative data in terms of N<sub>2</sub>O release are still lacking to-date. The total amount of N2O released by cattle is probably very small, because the gut is highly anoxic and this will favor the formation of NH<sub>3</sub>/NH<sub>4</sub><sup>+</sup> (Tiedje, 1988). Direct losses from animals themselves are likely to be much less than 10 g N<sub>2</sub>O-N/kg N excreted or taken up by the animal. Therefore, it is not included in the emission estimate.

# B. N<sub>2</sub>O emissions from animal waste management systems

The proportion of total N intake that is excreted and its partition between urine and feces is dependent on the type of animal, the intake of dry matter, and the N concentration of the diet (Whitehead, 1970). The retention of N in animal products (i.e. milk, meat, wool and eggs) generally ranges from about 5 to 20% of the total N intake. The remainder is excreted via dung and urine. For sheep and cattle, fecal excretion is usually about 8 g/kg of dry matter consumed, regardless of the N content of the feed (Haynes and Williams, 1993). The remainder of the N is excreted in the urine and as the N content of the diet increases, so does the proportion of N in the urine. In intensive animal production systems, where animal intake of N is high, more than half of the N is excreted as urine.

Production of  $N_2O$  during storage and treatment of animal wastes can occur via combined nitrification-denitrification of ammoniacal N contained in the wastes. The amount released depends on the system and duration of waste management. As fresh dung and slurry is highly anoxic and well-buffered with near neutral pH, one would expect  $N_2O$  production to increase with increasing aeration. Aeration initiates the nitrification-denitrification reactions, and hence makes release of  $N_2O$  possible. Unfortunately, there is not enough quantitative data to derive a relationship be-

FRACBURN	= 0.25 in developing countries; 0 in developed countries (kg N/kg crop-N)
FRACFUEL	= justified in-country estimate, or 0.0 kg N/kg N excreted
FRACGASF	= $0.1 \text{ kg NH}_3$ –N + NO <sub>x</sub> –N/kg of synthetic fertilizer N applied
FRACGASM	= $0.2 \text{ kg NH}_3$ –N + NO <sub>x</sub> –N/kg of N excreted by livestock
FRACGRAZ	= Tables 5,6 and Appendix 1
FRACLEACH	= 0.3 kg N/kg N of fertilizer or manure
FRACNCRBF	= 0.03 kg N/kg of dry biomass
FRACNCR0	= 0.015 kg N/kg of dry biomass
FRACNPR	= 0.16 kg N/kg of protein
FRACR	= 0.45  kg N/kg crop-N

tween the degree of aeration and  $N_2O$  emission from slurry during storage and treatment. There is a wide range in estimated losses, when expressed in g  $N_2O$  N per kg N in the waste. Losses from animal waste during storage range from < 0.1 g N for slurries to > 150 g N per kg N for pig waste in deep-litter stables (Groenestein et al., 1993; Kroeze, 1994; Sibbesen and Lind, 1993). Hence, different emission factors are needed for different animal waste management systems (Table 6). We use emission factors of 1, 20, and 5 g  $N_2O$ –N kg $^{-1}$  of N excreted for anaerobic lagoons and liquid systems, solid storage and drylot and for all other animal waste management systems, respectively. These emission factors should be revised as new data become available.

#### C. N<sub>2</sub>O from animal grazing

A brief summary of estimates of N2O emissions derived from dung and urine deposits of grazing animals is compiled in Table 4. The N2O emission is expressed as per cent of the N in urine and/or dung. Two types of studies may be distinguished. The first type focuses on emissions from a well-defined urine and/or dung patch. A control treatment is generally included, to facilitate the calculation of urine and dung derived emissions. The grazed grassland is the focus in the second type of studies. Grazing derived emissions can be obtained properly when a non-grazing treatment is included. For the purpose of this compilation we consider that grazing derived emissions are similar to 'dung and urine derived' emissions. This may not be completely true, because grazing animals have also other effects than deposition of dung and urine. For example, compaction of the soil by trampling may increase N<sub>2</sub>O emission from soil (Oenema et al., 1997).

The duration of the studies ranged from 1 week up to 2 years. Though the bulk of the N<sub>2</sub>O will be lost shortly after deposition in the field, significant amounts may still be released from the urine and dung even after a couple of weeks after deposition. Hence, short-term studies may under-estimate the total N2O losses from animal excrements (Van Cleemput et al., 1994; Velthof et al., 1996a). Grazing derived emissions range from 2 to 98 g N<sub>2</sub>O-N/ kg of N excreted (Table 4). The lower estimates are from well-drained unfertilized grassland soils in New Zealand (Carran et al., 1995). Large grazing derived emissions, induced by livestock N excretion, were obtained on drained peat soils in the Netherlands. These intensively managed grassland on peat soils have also a large background emission and also a large fertilizer derived emission (Velthof and Oenema, 1995; Velthof et al., 1996a).

Nearly all data pertain to temperate areas, with intensively managed grassland. The N contents of dung and especially urine are higher from these intensively managed grassland than from the less intensively managed (sub)tropical grasslands. The fraction of easily hydrolyzable N, i.e. urea and uric acid, is much smaller in dung and urine from animals fed with a low N content ration than from animals fed with a high N content ration. This difference will probably result in a different emissions factor. Unfortunately, data are lacking to sustain this hypothesis. Differences in climate, i.e. rainfall and temperature patterns, may also have a significant effect.

Nitrogen losses as  $N_2O$  are probably lower in arid and semiarid regions. Mosier and Parton (1985) found that during the course of a year in a semiarid grassland per kg of urea-N from simulated urine patches, only 6 g  $N_2O$ -N was emitted as  $N_2O$ . They did find in

Table 4. Emission of  $N_2O$  from animal dung and urine deposited in grassland; a compilation of published and unpublished data. The emitted amount of  $N_2O$  is expressed in % of the amount of N excreted by the grazing animal (see also Oenema et al., 1997

Country	Soil type	Treatment	Period	N <sub>2</sub> O Emission	Reference
United Kingdom	clay loam	urine	4 weeks	1-5	Monaghan and
					Barraclough (1993)
New Zealand	silt loam	urine	6 weeks	< 0.5	Sherlock and Goh (1983)
Germany	loess	urine	11 weeks	3.8	Flessa et al. (1996)
Germany	Loess	dung	11 weeks	0.5	Flessa et al. (1996)
The Netherlands	clay	urine	4 weeks	0.5	Velthof and Oenema
					(1994)
United Kingdom	clay loam	grazing	1 week	1.8	Velthof et al. (1996b)
The Netherlands	sand	grazing	32 weeks	1.0	Velthof and Oenema
					(1995)
The Netherlands	peat I	grazing	32 weeks	1.5	Velthof and Oenema
					(1995)
The Netherlands	peat II	grazing	32 weeks	7.7	Velthof and Oenema
					(1995)
Germany	loam	urine/dung	1 year	0.4-1.3	Poggeman et al. (1995)
The Netherlands	sand	grazing	2 years	1.5	Velthof et al. (1996a)
The Netherlands	clay	grazing	2 years	3.3	Velthof et al. (1996a)
The Netherlands	peat I	grazing	2 years	2.3	Velthof et al. (1996a)
The Netherlands	peat II	grazing	2 years	9.8	Velthof et al. (1996a)
New Zealand	silt loam	grazing	1 year	0.2-1.0	Carran et al. (1995)

later studies that  $N_2O$  emissions remained detectably higher 10 years after the urea had been applied to the semiarid shortgrass prairie (Mosier et al., 1991).

An overall reasonable average emission factor for animal waste excreted in pastures is 20 g  $N_2O-N/kg$  of N excreted. Note that this emission factor is almost twice as large as the emission factor for animal wastes spread on agricultural fields following manure storage. This difference is thought to be related to the high N content in the dung and urine patches.

Methodology for estimating  $N_2O$  from animal production

A simple methodology which takes into account the type and number of animals, the N excretion by the animals, and the Animal Waste Management System (AWMS), which includes free-range grazing is described below. Specific emission factors, expressed in per cent of N excreted are assigned to the animals and to the AWMS, representing direct emissions from the AWMS. Hence, the total emission from animal production in a country follows from:

 $N_2OANIMALS = \Sigma(N_2OAWMS)$ 

where

$$\begin{split} &N_2OAWMS = [N(T=1)*NEX(T=1)*AWMS(T=1)\\ *EF3(AWMS)] + ... + [N(T=TMAX)*NEX(T=TMAX)\\ *AWMS(T=TMAX)*EF3(AWMS)]\\ &and: \end{split}$$

AWMS(T) = fraction of NEX(T) that is managed in one of the different distinguished animal waste management systems for animals of type T in the country, Appendix 1

EF3(AWMS) =  $N_2O$  emission factor for an AWMS (kg  $N_2O$ –N/kg of NEX in AWMS), Table 6  $N_2O$ ANIMALS =  $N_2O$  emissions from animal production and pastures (kg N yr  $^{-1}$ ) from a country  $N_2O$ AWMS =  $N_2O$  emissions from Animal Waste Management Systems in the country (kg N yr  $^{-1}$ ) N(T) = number of animals of type T in the country NEX(T) = N excretion of animals of type T in the country, Table 5

T =type of distinguished animals

TMAX = last type of animal distinguished in the country

#### Nitrogen excretion

General statistics about animal numbers are provided by FAO in their Production Yearbooks (FAO, 1956–

*Table 5.* Default values for N excretion per head of animal per region (kg N animal $^{-1}$  yr $^{-1}$ )

Region	Type of animals					
	Non-dairy cattle	Dairy cattle	Poultry	Sheep	Swine	Other animals
North America	70	100	0.6	16	20	25
Western Europe	70	100	0.6	20	20	25
Eastern Europe	50	70	0.6	16	20	25
Oceania	60	80	0.6	20	16	25
Latin America	40	70	0.6	12	16	40
Africa	40	60	0.6	12	16	40
Near East &						
Mediterranean	50	70	0.6	12	16	40
Asia & Far East	40	60	0.6	12	16	40

1994) and detailed information is available for many countries. Default values for N excretion for each animal type and region are provided in Table 5, which was compiled on the basis of data provided by ECETOC (1994), and references therein, Vetter et al. (1988), Steffens and Vetter (1990). There are still uncertainties in the values listed in Table 5. Estimates for cattle and swine may be too high. Hence, these estimates (default values) need further attention. Countries may chose to use N excretion data from the Ammonia Expert Panel of the UN-ECE task force on emission inventories.

### Animal waste management systems

The types of Animal Waste Management Systems (AWMS) distinguished by Safley et al. (1992) and their compilations for a large number of countries are proposed for this methodology (Appendix 1). Descriptions of these management systems can be found in Vol. 3 of the IPCC Guidelines, in Table 4–8 and Table 4–6 (IPCC, 1995b). The AWMS is an important regulating factor in N<sub>2</sub>O emission from animal wastes during storage and treatment. The data provided per country in Safley et al. (1992) could be used for estimating N<sub>2</sub>O emissions from animal wastes. Significant differences in emission factors are expected between some of the AWMS.

The class 'used for fuel' is not included here as a source of  $N_2O$ , because this possible source of  $N_2O$  is considered an energy-related emission. Nevertheless, countries should estimate the amount of manure N that is used as fuel, because that amount is not applied to soils (see Appendix 1).

Default emission factors (EF3) for the different AWMS are shown in Table 6. These factors were

Table 6. Tentative default values for  $N_2O$  emission factors from animal waste per Animal Waste Management System, in g  $N_2O$ -N per kg N excreted

Animal Waste Management System <sup>a</sup>	Emission factor EF3
Anaerobic lagoons	1 (0 - 2)*
Liquid Systems	1 (0 - 1)
Daily spread	$0.0  (\text{no range})^b$
Solid storage & drylot	20(5-30)
Pasture range & paddock (grazing)	20 (5 – 30)
Used as fuel	not included
Other systems	5 (no range)
*indicates range	

a See Appendix 1

derived on the basis of a very limited amount of information. Uniform factors for all over the world are proposed. This may be incorrect, as temperature and moisture may have positive effects on the size of the processes and, hence, on losses. However, animal production systems in warmer regions, are low-intensity systems and generally have less easily hydrolyzable N in the excretions, therefore, less N<sub>2</sub>O production. These counteracting effects suggest that a uniform factor for all regions would seem appropriate.

# 4. Indirect N<sub>2</sub>O emissions from N used in agriculture

Overview of sources for indirect N2O production

The pathways for synthetic fertilizer and manure nitrogen input that give rise to indirect emissions are considered to be:

- A. Volatilization and subsequent atmospheric deposition of NH<sub>3</sub> and NO<sub>x</sub>
- B. Nitrogen leaching and runoff
- C. Human consumption of crops followed by municipal sewage treatment
- D. Formation of N<sub>2</sub>O in the atmosphere from NH<sub>3</sub>
- E. Food processing

Of these, methodologies for estimating  $N_2O$  emissions from A–C are proposed. At present, insufficient information is available to estimate emissions from D and E.

#### A. Atmospheric deposition of $NO_x$ and $NH_3$

Atmospheric deposition of nitrogen compounds such as nitrogen oxides  $(NO_x)$  and ammonium (from  $NH_3$ )

b Assumes no storage of manure; emissions after use as fertilizer are included in direct soil emissions.

fertilize soils and surface waters and as such enhance biogenic  $N_2O$  formation. Indeed, Brumme and Beese (1992) showed that after two decades of atmospheric deposition of acidifying compounds (ammonium and sulphuric acids),  $N_2O$  emissions from German forest soils were enhanced by up to a factor of 5. Reported rates of  $N_2O$  emissions are between 0.2 and 1.6% of the amount of N deposited onto soils (Bowden et al., 1991; Brumme and Beese, 1992; Kasimir, personal communication). This is within the range of emission factors suggested for synthetic fertilizers. We therefore calculate  $N_2O-N$  emissions as 1% of  $NO_x-N$  and  $NH_3-N$  emission in a country (EF4, Table 2).

Animal manure (dung + urine) is one of the most important sources of atmospheric NH3. According to Van der Hoek (1994), up to 50% of the mineral N in animal manure (i.e. about 25% of total N) may be lost shortly as NH<sub>3</sub> after application to soil. He also shows that this percentage depends considerably on the application technique used. Schimel et al. (1986) assumed that, as a minimum estimate, 20% of manure nitrogen applied to soils is volatilized as NH<sub>3</sub> soon after application. Although climate and fertilizer type (e.g. Urea or ammonium sulfate) may influence ammonia volatilization, we use the following default for NH<sub>3</sub> and NO<sub>x</sub> volatilization: 0.1 kg N/ kg synthetic fertilizer N applied to soils and 0.2 kg N/ kg of N excreted by livestock (FRACGASF and FRACGASM, Table 3). These N<sub>2</sub>O–N emissions are to be calculated from a country's NO<sub>x</sub> and NH<sub>3</sub> emissions and N transported in leaching and runoff, so that all N2O formed as a result of  $NO_x$  and  $NH_3$  emissions and leaching and runoff in country Z are assigned to country Z, even if the actual N2O formation takes place in another country. This implies that NO<sub>x</sub> and NH<sub>3</sub> and N from leaching and runoff imported into a country is not included in the calculations.

#### B. Leaching and runoff

A considerable amount of fertilizer N is lost from agricultural soils through leaching and runoff. The leached/runoff N enters groundwater, riparian areas and wetlands, rivers and eventually the coastal ocean. In many world areas it is one of the most important inputs of nitrogen to those systems. A WHO/UNEP report (1989) showed that over 10% of European rivers had a nitrate content ranging from 9 to 25 mg nitrate-N/L; in the US 10 mg nitrate-N/L is considered unfit for human consumption. Mineral N in ground water and surface waters enhances biogenic

production of  $N_2O$  as the N undergoes nitrification and denitrification.

The amount of nitrogen lost to leaching and surface runoff may differ considerably between watersheds (Kroeze and Seitzinger, this volume). We therefore estimate that the fraction of the fertilizer and manure N lost to leaching and surface runoff (FRACLEACH) may range from 0.1–0.8. For this purpose total N excretion (NEX) is used in order to include manure produced during grazing:

# NLEACH = [NFERT + NEX]\*FRACLEACH

The sum of the emission of  $N_2O$  due to NLEACH in: 1) groundwater and surface drainage (EF5-g), 2) rivers (EF5-r), and 3) coastal marine areas (EF5-e) is calculated to obtain the  $N_2O$  emission factor (EF5) for NLEACH. Although not specified, the total amount of N eventually denitrified remains the same but some is denitrified in riparian area and wetlands before the N reaches the ocean. In future assessment methodologies, a separate emission factor should be used for each of these three environments.

# Background information for developing emission factor (EF5) for NLEACH.

Groundwater and surface drainage. Supersaturated concentrations of N2O in groundwater and in surface water draining agricultural lands may occur due to leaching of N2O from the soil towards drainage and groundwater, or production during nitrification and/or denitrification of fertilizer N in the groundwater or drainage ditches. Mineral N in groundwater or drainage water is primarily in the form of NO<sub>3</sub>-N. While the range of N<sub>2</sub>O concentrations reported is large, there is some relationship between the concentration of N<sub>2</sub>O-N and NO<sub>3</sub>-N in groundwater and agricultural drainage water. The ratio of N<sub>2</sub>O-N to NO<sub>3</sub>-N concentration in groundwater and agricultural drainage water at over 25 locations in urban, agricultural and woodland areas in Japan, Israel and the United States ranged from 0.0001 to 0.06 (Dowdell et al., 1979; Minami and Fukuski, 1984; Ronen et al., 1988; Minami and Oshawa, 1990; Ueda et al., 1991; Ueda et al., 1993). The ratio of N<sub>2</sub>O-N to NO<sub>3</sub>-N in agricultural drainage ditches and groundwater under agricultural fields ranged from approximately 0.0003 to 0.06. The ratio of N<sub>2</sub>O-N to NO<sub>3</sub>-N in agricultural drainage ditches were generally lower (0.003 or less) than ratios in agricultural groundwater. Rapid loss of N<sub>2</sub>O to the atmosphere may account for the generally lower ratios in drainage ditch water. The ratio of N<sub>2</sub>O–N to NO<sub>3</sub>–N in agricultural groundwater was generally between 0.003 and 0.06, with values between 0.007 and 0.02 common. Assuming that all NLEACH is in the form of NO<sub>3</sub>, we recommend a default emission factor of 0.015 (EF5-g) for N<sub>2</sub>O from NLEACH in groundwater and drainage ditches, with a range of 0.003 to 0.06. The amount of N<sub>2</sub>O emitted from groundwater (by upward diffusion or following entry of groundwater into surface water through rivers, irrigation, and drinking water) and agricultural drainage water is then estimated as:

#### NLEACH\* EF5-g

where EF5-g = 0.015 kg N<sub>2</sub>O–N/ kg NLEACH, assuming that all N<sub>2</sub>O produced in a particular year is emitted during that year.

*Rivers.* Once NLEACH from groundwater and surface water enters rivers, additional N<sub>2</sub>O is produced associated with nitrification and denitrification of NLEACH (Seitzinger and Kroeze, in prep.).

Nitrification: N2O can be produced during nitrification of NLEACH in rivers. While most of the NLEACH enters rivers as nitrate, algae and aquatic plants can assimilate the nitrate into organic matter, which is released as ammonia, following decomposition of that organic matter. Ammonia in rivers is rapidly nitrified (Lipschultz et al., 1986). The NLEACH entering rivers nitrifies on average 0.5 - 3times during river transport. We assume for our default methodology that all NLEACH entering rivers is nitrified once during river transport. The  $N_2O$  yield (moles N<sub>2</sub>O-N per mol of NO<sub>3</sub>-N) during nitrification is generally between 0.002 and 0.003 at atmospheric oxygen levels (0.2 atm partial pressure) (Watson, 1965; Watson and Remsen, 1969; Goreau et al., 1980), enhanced yields of N<sub>2</sub>O are found at reduced O<sub>2</sub> concentrations (Goreau et al., 1980). We use an N2O yield of 0.005 for nitrification.

Denitrification: During river transport a considerable amount of nitrogen is lost via denitrification. A wide range of denitrification rates has been measured in riverine sediments; rates are generally lowest in unpolluted streams (Duff et al., 1984) with highest rates in polluted rivers/streams (Robinson et al., 1979; Cooper and Cooke, 1984; Seitzinger, 1988, 1990; Christensen and Sorensen, 1988; Christensen et al., 1989). Factors likely to affect the fraction of N removed by denitrification include length and depth of the river, flow rate, water residence time, oxygen content, organic content of sediments, and season. In

a number of rivers denitrification removed 50% of the N inputs, even over short sections (Kaushik and Robinson, 1976; Hill, 1979, 1981, 1983; van Kessel, 1977; Swank and Caskey, 1982). Here, we assume that denitrification removes 50% of NLEACH inputs to rivers.  $N_2O$  associated with denitrification (Jorgensen et al., 1984) is released from river sediments. The ratio of  $N_2O:N_2$  emitted from river sediments is generally within the range 0.001-0.005, although in heavily polluted sediments yields up to 6% have been observed (Seitzinger, 1988). A constant ratio of 0.005 for  $N_2O-N$  emission to denitrification ( $N_2-N$  production) in rivers is suggested.

In summary, the emission factor for NLEACH in rivers due to nitrification and denitrification [EF5-r] is thus equal to 0.005\*NLEACH [for nitrification] plus 0.005\*(NLEACH/2) [for denitrification], or 0.0075\*NLEACH. Therefore,  $N_2O-N$  produced from NLEACH during river transport = NLEACH\*(EF5-r), where EF5-r = 0.0075.

Estuaries. Rivers are the major conduit for N transport to the coastal ocean (estuaries). As discussed above, half of NLEACH is assumed to be removed by denitrification in rivers in the form of  $N_2$  and  $N_2O$ . The remaining 50% of NLEACH is discharged by rivers to estuaries. N inputs to estuaries can undergo nitrification and denitrification, with associated  $N_2O$  production.

Nitrification: Nitrification rates in estuaries generally range from 0–22  $\mu$ mol l<sup>-1</sup> d<sup>-1</sup> (Berounsky and Nixon, 1993). Estuarine nitrification rates are affected by a number of factors such as ammonia concentrations, temperature (Berounsky and Nixon, 1985, 1993), oxygen (Helder and DeVries, 1983), suspended particulate matter (Helder and DeVries, 1983; Owens, 1986), and light (Horrigan and Springer, 1990). However, no predictive factor has been developed to estimate pelagic nitrification rates across a range of estuaries. In Narragansett Bay (USA), approximately half of the river inputs of inorganic N to the Bay were nitrified in the bay (Beroundsky and Nixon, 1993; Seitzinger and Kroeze, in prep.). Herein, we assume that half of the rivers inputs of NLEACH are nitrified again in estuaries, and that the ratio of N<sub>2</sub>O-N to NO<sub>3</sub>-N produced is 0.005, as discussed above for rivers.

Denitrification: A relatively good relationship has been found between denitrification and inorganic N inputs to estuaries from rivers (Kemp et al., 1990; Jenkins and Kemp, 1984; Jensen et al., 1984, 1988;

Smith et al., 1985) which is equivalent to a relatively constant percentage (50%) of inorganic N inputs to a variety of estuaries (Seitzinger, 1988). Those estuaries vary in a number of characteristics including N loading rates (25 to  $516 \times 10^{-6}$  mol N m<sup>-2</sup> h<sup>-1</sup>), extent of intertidal area (< 1% to 50%), and latitude (subtropical to subarctic). We use 50% of the NLEACH that is carried to estuaries by rivers is denitrified, and the ratio of N<sub>2</sub>O–N to denitrification (N<sub>2</sub>–N) emitted is 0.005, as discussed above for rivers. NLEACH that enters estuaries but is not denitrified, is either buried in the sediments as organic N or exported to the continental shelf region where additional N<sub>2</sub>O can be produced. Nitrous oxide production associated with this fraction of NLEACH is not accounted for in this methodology.

In summary, the Phase II methodology assumes the following: 1) half of the NLEACH is transported to estuaries by rivers, 2) half of the NLEACH in estuaries is nitrified again in the estuary with a ratio of  $N_2O-N$  to  $NO_3-N$  of 0.005, and 3) half of the NLEACH in estuaries is denitrified in the estuary with a  $N_2O-N$  to  $N_2-N$  ratio of 0.005. Therefore,  $N_2O-N$  produced from NLEACH in estuaries = NLEACH\*(EF5-e) where EF5-e = 0.0025.

The combined emission factor [EF5] for  $N_2O$  due to NLEACH in: 1) groundwater and surface drainage (EF5-g = 0.015 kg  $N_2O$ –N per kg NLEACH), 2) rivers (EF5-r = 0.0075 kg  $N_2O$ –N per kg NLEACH), and 3) coastal marine areas (EF5-e = 0.0025 kg  $N_2O$ –N per kg NLEACH) is 0.025 (EF5). Therefore:

# $\begin{aligned} NLEACH &= [NFERT + NEX]*FRACLEACH \\ N_2O(L) &= NLEACH*EF5 \end{aligned}$

where the default values are FRACLEACH=0.3 kg N/kg N input to soils and EF5=0.025 kg  $N_2O$ -N/kg NLEACH.

# C. Human consumption followed by municipal sewage treatment

Consumption of foodstuffs by humans results in the production of sewage. Sewage can be disposed of directly on land (night-soil or spray irrigation) or discharged into a water source (e.g. rivers and estuaries). Before disposal on land or into water, it also can be processed in septic systems or wastewater treatment facilities. During all of these stages, N<sub>2</sub>O can be produced during nitrification and denitrification of sewage nitrogen. In the 1997 Guidelines (IPCC, 1997) N<sub>2</sub>O emissions from sewage will be reported as waste emissions rather than under agricultural sources.

Sewage N (NSEWAGE) production can be estimated from FAO per capita protein consumption data (PROTEIN) and human population counts (NR-PEOPLE), assuming that N constitutes about 16% by weight of protein (FRACNPR, Table 3): Sewage N production can be estimated from FAO per capita protein consumption data (PROTEIN) and human population counts (NRPEOPLE). Protein consumption may vary by a factor of 2 between countries, e.g., Americans and Indians consume 110 and 55 g protein/person/day, respectively.

#### NSEWAGE = PROTEIN\*FRACNPR\*NRPEOPLE

Nitrous oxide emissions resulting from sewage nitrogen are estimated following: 1) land disposal or wastewater treatment of sewage, and 2) input of sewage N to rivers and estuaries.

Disposal or wastewater treatment of sewage. studies were found quantifying nitrous oxide emissions from land disposal of sewage, although supersaturated concentrations of N2O in groundwater under cultivated land irrigated with sewage effluent have been reported (Ronen et al., 1988). A few studies have documented N2O emission associated with wastewater treatment operations (e.g. Benckiser et al., 1996; Czepiel et al., 1995; Debruyn et al., 1994; Hemond and Duran, 1989; Hanaki et al., 1992; Hong et al., 1993; Velthof and Oenema, 1993). All studies reported low rates of N<sub>2</sub>O emission. For example, nitrous oxide emissions from a secondary treatment wastewater facility in New Hampshire (USA) were approximately 0.0006 g N2O-N/g sewage N, assuming 3.2 kg sewage N are produced person<sup>-1</sup> yr<sup>-1</sup> (Czepiel et al., 1995). Velthof and Oenema (1993) found N<sub>2</sub>O losses of 22 g d<sup>-1</sup> in a vented closed waste water treatment facility that had a daily input of 900 kg N, suggesting that N<sub>2</sub>O losses were 0.00005 g/g N entering the system. Additional N2O released to the atmosphere following discharge of supersaturated effluent to the environment is also low (0.0007 g  $N_2O-N/g$  sewage N) (Hemond and Duran, 1989).

For the our estimates  $N_2O$  associated with sewage treatment and land disposal is assumed to be negligible. This is based on the low emission rates of  $N_2O$  reported for operating wastewater treatment facilities (Hemond and Duran, 1989; Czepiel et al., 1995; Velthof and Oenema, unpubl. results), and the lack of information on  $N_2O$  production from land disposal of human sewage. This assumption should be reviewed in the future, as new data become available.

Rivers and estuaries. N<sub>2</sub>O is produced in rivers and estuaries following nitrification and denitrification of sewage N inputs (Seitzinger and Kroeze, in prep.). The sewage nitrogen can be discharged directly to aquatic environments (e.g. rivers, estuaries) or enter aquatic environments following leaching from terrestrially disposed sewage. Here, it is assumed that minimal removal of sewage nitrogen occurs during land disposal or sewage treatment, and that all sewage nitrogen enters rivers and/or estuaries. This latter assumption should be reviewed in the future, as more data become available.

Nitrous oxide emissions in rivers and estuaries due to nitrification and denitrification of sewage N are estimated using the same assumptions used for mineral N leached to rivers and estuaries (Section B). These assumptions result in emission coefficients of EF6-r = 0.0075 g N<sub>2</sub>O–N/g NSEWAGE (rivers) and EF6-e = 0.0025 g N<sub>2</sub>O–N/g NSEWAGE (estuaries). The sum of N<sub>2</sub>O emissions in rivers (0.0075 \* NSEWAGE) and estuaries (0.0025 \* NSEWAGE) associated with nitrification and denitrification of NSEWAGE is calculated as:

 $\label{eq:N2O} N_2O(S) = NSEWAGE * EF6$  where EF6 = 0.01 g N\_2O–N/ g NSEWAGE

D. Formation of  $N_2O$  in the atmosphere from  $NH_3$ Dentener and Crutzen (1994) proposed that oxidation of NH<sub>3</sub> and subsequent reaction of the intermediate NH<sub>2</sub> radical with NO<sub>2</sub> could lead to a production of  $0.6 + 0.3 \text{ Tg N}_2\text{O-N yr}^{-1}$ . They parameterized natural NH<sub>3</sub> emissions from vegetation using a highly uncertain NH<sub>3</sub> canopy compensation point (the atmospheric concentration above which plants assimilate and below which they emit NH<sub>3</sub>). Without considering this compensation point, N2O production was reduced by 55%. Other sources of NH<sub>3</sub> in the tropics include animal waste decomposition (both from wild and domestic animals), fertilizer application and biomass burning emissions. About half of the atmospheric N<sub>2</sub>O production may be associated with agricultural nitrogen, amounting to about 0.25 Tg  $N_2O-N$  yr<sup>-1</sup>. Due to the high uncertainty of this estimate (ca. 100%), we have not included this potentially important source in our agricultural N2O emissions inventory. More measurements on the co-occurance of high NH<sub>3</sub>, NO<sub>2</sub> and OH concentrations in the tropics are needed to provide more insight in the photochemical production of  $N_2O$ .

E. Food processing operations

Some food processing operations are sources of  $N_2O$ . A fraction of the edible crop harvest is not consumed by people and enters the waste stream, for instance when it is landfilled, composted, burned or fed to animals. At this point, there are no data to calculate the size of this  $N_2O$  source.

Methodology for estimating indirect  $N_2O$ 

Based on the above, we propose the following methodology for calculating a country's indirect  $N_2O$  emissions (kg N yr<sup>-1</sup>):

$$N_2OINDIRECT = N_2O(G) + N_2O(L) + N_2O(S)$$

where

 $N_2O(G) = N_2O$  produced from atmospheric deposition of  $NO_x$  and  $NH_3$  (kg N yr<sup>-1</sup>)

= (NFERT \* FRACGASF + NEX \* FRACGASM) \* FF4

 $N_2O(L) = N_2O$  produced from N leaching and runoff (kg N yr<sup>-1</sup>)

= NLEACH \* EF5

 $N_2O(S) = N_2O$  produced from human sewage (kg  $N \text{ yr}^{-1}$ )

= NSEWAGE \* EF6

EF4 = emission factor for atmospheric deposition (kg  $N_2O-N/kg$   $NH_3-N$  and  $NO_x-N$  emitted); Table 2.

EF5 = emission factor for leaching/runoff (kg  $N_2O-N/kg\ N$  leaching/runoff); Table 2.

EF6 = emission factor for sewage treatment (kg  $N_2O-N/kg$  sewage-N produced); Table 2.

FRACGASF = fraction of synthetic fertilizer N applied to soils that volatilizes as  $NH_3$  and  $NO_x$  (kg  $NH_3$ –N and  $NO_x$ –N/kg of N input); Table 3.

FRACGASM = fraction of livestock N excretion that volatilizes as NH<sub>3</sub> and NO<sub>x</sub> (kg NH<sub>3</sub>–N and NO<sub>x</sub>–N/kg of N excreted); Table 3.

FRACNPR = fraction of N in protein (kg N/kg of protein); Table 3.

FRACLEACH = fraction of N input to soils that is lost through leaching and runoff (kg N/kg of N applied); Table 3.

NLEACH = N leaching in country (kg N yr<sup>-1</sup>)= (NFERT + NEX \* 0.8) \* FRACLEACH

NRPEOPLE = number of people in country. FAO data. NSEWAGE = PROTEIN \* NRPEOPLE \* FRACNPR PROTEIN = annual per capita protein consumption in country (the global average is 25.6 kg protein person $^{-1}$  yr $^{-1}$ ).

# 5. Application of methodology for total $N_2O$ emissions to the world

Total emissions of  $N_2O$  from agriculture in 1989 can be calculated as the sum of direct emissions, emissions from animal production and indirect emissions. Thus, Total  $N_2O-N$  emissions from the world (kg  $N_2O-N$  yr<sup>-1</sup>) are:

 $N_2O = N_2ODIRECT + N_2OANIMALS + N_2OINDIRECT$ 

### STEP I. Input data collection

#### Input data:

- 1. Worldwide use of synthetic fertilizer NFERT = 79 Tg N yr<sup>-1</sup> (FAO, 1990b).
- 2. Number of livestock in world for the following categories: non-dairy cattle, dairy cattle, poultry, sheep, swine and other animals: see FAO data. Total N excretion by livestock NEX = 138 Tg N yr<sup>-1</sup> (FAO data; IPCC, 1995b).
- 3. Dry biomass production of pulses and soybean (CROPBF) assumes 15% water content = 191 Tg x 0.85 = 162 Tg yr<sup>-1</sup> (FAO, 1990b).
- 4. Dry biomass production of other crops (CROP0), assuming 15% water content = 1870 Tg x 0.85 = 1590 Tg/y (FAO, 1990b).
- 5. Area of organic soils (Histosols) (FOS, ha) = 286 x 10<sup>5</sup> in tropics, 1459 x 10<sup>5</sup> in rest of world (Eswaren et al., 1993), assume 10% of Histosols are cultivated in nontropical areas and 10% in tropics.
- 6. Average annual per capita protein intake in world  $(PROTEIN) = 25.6 \text{ kg person}^{-1} \text{ yr}^{-1})$  (FAO).
- 7. Number of people in world (NRPEOPLE) = 5.3 billion (FAO).

### STEP II: Calculation of fertilizer use (FSN)

Total synthetic fertilizer use excluding emissions of  $NH_3$  and  $NO_x$  (FSN):

 $FSN = NFERT * (1-FRACGASF) = 79 * (1-0.1) = 71.1 Tg N yr^{-1}.$ 

STEP III: Calculation of livestock N excretion (NEX and FAW)

Total animal waste N used as fertilizer (FAW): FAW = (NEX \* (1- (FRACFUEL + FRACGRAZ + FRACGASM)) = 138 \* (1-(0.1 + 0.34 + 0.2) = 49.7 Tg N yr $^{-1}$  STEP IV. Calculation of N input from N-fixing crops (FBN)

N fixed by N-fixing crops (FBN): FBN = 2 \* CROPBF \* FRACNCRBF = 2 \* 162 \* 0.03=  $9.7 Tg N yr^{-1}$ 

STEP V. Calculation of N input from crop residues (FCR)

It is assumed that, worldwide, 10% of the crop residue is burned (FRACBURN = 0.1)

Crop residue returned to soils (FCR):  $FCR = 2 * [CROP0 * FRACNCR0 + CROPBF * FRACNCRBF] * (1-FRACR) * (1-FRACBURN) = 2 * [1590 * 0.015 + 162 * 0.03] * 0.55 * 0.9 = 28.4 Tg N yr^{-1}$ 

# STEP VI. Calculating direct N<sub>2</sub>O emissions

Direct N<sub>2</sub>O emissions from agricultural soils: N<sub>2</sub>ODIRECT (Tg N yr<sup>-1</sup>) = [(FSN + FAW + FBN + FCR) \* EF1] + (Area of tropical cultivated Histisols \* 10 kg N ha-1 yr-1) + (Area of cultivated Histosols in rest of world \* 5 kg N<sub>2</sub>O–N ha-1 yr-1)] = [(71.1 + 49.7 + 9.7 + 28.4) \* 0.0125] + [0.03 + 0.07] = 2.0 + 0.1 = 2.1 Tg N yr<sup>-1</sup>.

STEP VII Calculating  $N_2O$  emissions from animal production

 $N_2O$  emissions from animal production ( $N_2OANIMALS$  in Tg N yr<sup>-1</sup>) as:

 $\begin{array}{lll} N_2O & ANIMALS & = & N_2O(T,AWMS) & = & [N(T=1)\\ *NEX(T=1)*AWMS(T=1)*EF3(AWMS)] + & ... & + \\ [N(T=TMAX)*NEX(T=TMAX)*AWMS(T=TMAX)\\ *EF3(AWMS)] & = & 2.1\ Tg\ N\ yr^{-1}. \end{array}$ 

# STEP VIII. Calculating indirect emissions

Indirect  $N_2O$  emissions (kg yr $^{-1}$ ) can now be calculated (1) (atmospheric deposition), (2) (leaching and runoff) and (3) (human sewage) as:  $N_2OINDIRECT = N_2O(G) + N_2O(L) + N_2O(S) = [(NFERT*FRACGASF+NEX*FRACGASM)*EF4] + [(NFERT + NEX *FRACLEACH) * EF5] + [(PROTEIN*NRPEOPLE*FRACNPR) * EF6] = [0.3 + 1.6 + 0.2] = 2.1 Tg N yr<math>^{-1}$ 

# STEP IX. Total N<sub>2</sub>O emissions from agriculture

Total emissions of N<sub>2</sub>O from agriculture can be calculated as the sum of direct emissions, emissions from

Table 7. Global  $N_2O$  emissions from agricultural soils calculated with the PHASE II methodology (Tg N yr $^{-1}$ )

Direct soil emissions	
- synthetic fertilizer	0.9(0.18-1.6)*
- animal waste	0.6(0.12-1.1)
- biological N <sub>2</sub> fixation	0.1(0.02-0.2)
- crop residue	0.4(0.07-0.7)
- cultivated Histosols	0.1(0.02-0.2)
- total	2.1(0.4-3.8)
Animal production	
- animal waste management systems	2.1(0.6-3.1)
Indirect emissions	
- atmospheric deposition	0.3(0.06-0.6)
- nitrogen leaching and runoff	1.6(0.13-7.7)
- human sewage	0.2(0.04-2.6)
- total	2.1(0.23-11.9)
Total	6.3(1.2-17.9)

 $<sup>^{\</sup>it a}$  Values in parentheses indicate estimated range which is derived from emission factor range in Table 2.

animal production and indirect emissions. Total  $N_2O-N$  emissions are calculated to be 6.3 Tg N yr $^{-1}$  (Table 7).

### 6. Future needs for methodology development

The methodology for country-based N2O described above is a rough, generalized approach which treats all agricultural systems as being the same under all climates, in all soils, in all crops and in all management systems. This clearly provides uncertainties in inventory calculations. We do feel, however, that the ranges of conversion factors provided cover much of the potential N<sub>2</sub>O emissions from each country, whatever climate, soils and set of crops is involved. Some recent studies in temperate (e.g. Thornton and Valente, 1996) and tropical (Veldkamp and Keller, 1997) show very high direct N<sub>2</sub>O emissions while other studies (Corrie et al., 1996; Flessa et al., 1995; Wagner-Riddle and Thurtel, this issue) demonstrate that significant N<sub>2</sub>O emissions commonly occur during thaw periods in early spring and winter or through snow-covered agricultural soils (Van Bochove et al., 1996). These recent studies indicate that many of the published direct estimates of N2O emissions from agricultural fields may be further underestimated because they did not account for cold season N2O emissions, which can be substantial.

To make significant improvement in inventory methodologies for  $N_2O$ , we felt that the next step is

to utilize process based models to produce country inventories, for direct emissions from agricultural soils (e.g. Li et al., 1992; Potter et al., 1996; Parton et al., 1996), appropriate animal management models for N<sub>2</sub>O from animal production, simulation models which more effectively represent N transformations in aquatic systems, including riparian areas, wetlands, rivers estuaries, continental shelves and the deep ocean (e.g. Seitzinger and Kroeze, submitted).

The soil C and N cycles are tightly integrated and we think that both C and N should be considered together so that various aspects of the C and N cycle and CO2 and N2O production can be more accurately defined. For example, the amount of N leached from agricultural fields represents a very large component of the global N<sub>2</sub>O production according to the methodology presented here. The accuracy of the N leaching fraction prediction is closely tied to C turnover in the soil as it controls N mineralization and immobilization. The turnover and retention of N in all soils is intimately linked with the C cycle. Conversely, C retention in soils is directly tied to mineral N availability. These models must, however, include adequate flexibility to predict cold soil emissions as well as emissions under tropical conditions.

Testing of the validity and accuracy of any inventory methodology needs to be conducted. Van Aardenne (1996) conducted an uncertainty and sensitivity analysis on the methodology presented in this paper. Using a Monte Carlo sampling technique and regression and correlation analyses he simulated the output of the methodology and determined the total uncertainty of the simulated model output and uncertainty contribution of 14 model parameters used for calculating emissions from soils, grazing animals, and indirect emissions. He found a standard deviation of 20% from calculated mean N2O emission values using the 1990 Dutch N values as a test situation. He discerned that the parameters EF1 (emission factor for direct soil emissions) and the FLH (expression of the fraction of N input to soils that is lost through leaching and runoff) were the largest contributors to the overall methodology uncertainty. This rather low estimate of error for the methodology is surprisingly small and suggests the need for further error analyses using country data that include a variety of climate and agricultural systems.

There are additional issues that should be addressed in the overall effort of estimating greenhouse gas exchange withing agricultural systems. These issues include: (1) development of methodologies that represent the effect of cropping system, soil, and climate on N<sub>2</sub>O budgets which account for N<sub>2</sub>O emissions per unit of food produced in various agricultural systems; (2) including soil methane oxidation in national budgets (without the soil sink component atmospheric methane concentrations would be increasing about two times faster than the increase rate observed in the 1980s) (Ojima et al., 1993); (3) including the impact of NO<sub>x</sub> emissions from agricultural soils on local and regional atmospheric oxidants and ozone concentration; (4) determining the impact of C and N losses and retention on system sustainability; (5) considering mitigation methodologies to decrease CO<sub>2</sub> and N<sub>2</sub>O emissions from agriculture and to improve the soil sink capacity for CH4; and (6) investigating errors that may arise as a result of aggregating field scale data to the national level. We assume that the methods used are applicable to small and large countries. However, there can be effects of 'scaling' for which there is no quantification.

#### 7. Summary

This paper presents the development of a new method for calculating national emissions of  $N_2O$  from agricultural soils. This  $N_2O$  method is a revision of the 1995 IPCC Guidelines for National Greenhouse Gas Inventories (IPCC, 1995b). The  $N_2O$  methodology was developed a group of experts, in which approximately 32 people from 18 countries participated.

The methodology includes more sources of  $N_2O$  and more explicit information about emission factors to be used than the Phase I IPCC Guidelines (IPCC, 1995). Three sources of  $N_2O$  are distinguished: (i) direct emissions from agricultural soils, (ii) emissions from animal production systems, and (iii)  $N_2O$  emissions indirectly induced by agricultural activities. The methodology is a simple approach which requires only input data that are available from FAO databases.

Our methodology attempts to relate N<sub>2</sub>O emissions to the agricultural nitrogen cycle and to systems into which N is transported once it leaves agricultural systems (Figure 1). Anthropogenic input into agricultural systems include synthetic fertilizer, N from animal wastes, N from increased biological N-fixation, N from crop residue returned to the field and N derived from cultivation of mineral and organic soils through enhanced organic matter mineralization. Nitrous oxide may be produced and emitted directly in agricultural fields, animal confinements or pastoral

systems. Part of the N applied to agricultural soils or from animal production systems is transported from agricultural systems into ground and surface waters through surface runoff, N leaching, consumption by humans and introduction into sewage systems which transport the N ultimately into surface water. Additional  $N_2O$  is produced during transport and residence in aquatic systems. Ammonia and oxides of N ( $NO_x$ ) are also emitted from agricultural systems and may be transported off-site and serve to fertilize other systems which leads to enhanced production of  $N_2O$ .

The methodology has a number of limitations because agricultural systems are considered as being the same throughout the world, and the methodology does not take into account different crops, soils and climates which are known to regulate  $N_2O$  production. It is likely that direct field emissions during the winter time from temperate soils as well as some tropical situations are underestimated. These factors are not considered because limited data are available to provide appropriate emission factors. The method also uses a linear extrapolation between  $N_2O$  emissions and fertilizer N application and in the indirect emissions section does not account for the probable lag time between N input and ultimate production of  $N_2O$  as a result of this N input into agricultural soils.

We estimated global N<sub>2</sub>O-N emissions for the year 1989, using midpoint emission factors from our methodology and the FAO data (FAO, 1990a,b) (Table 7). Direct emissions from agricultural soils totaled 2.1 Tg N, direct emissions from animal production totaled 2.1 Tg N and indirect emissions resulting from agricultural N input into the atmosphere and aquatic systems totaled 2.1 Tg N<sub>2</sub>O–N. These estimates show that each of the three components of agriculture considered contribute about the same amount of N2O to the global atmospheric budget. When looking at previous estimates (Table 1), the N2O input to the atmosphere from agricultural production as a whole has apparently been previously underestimated. We found that the total global N<sub>2</sub>O budget is reasonably in balance if we use the N2O emission estimate for agricultural soils calculated by the methodology presented in this paper (Table 1). The balance is tenuous, however, because of the large uncertainty in our estimate of global N<sub>2</sub>O from agriculture (Table 7) and the proportionation of natural and anthropogenic sources which needs to be reconciled. There are a few potential overlaps in sections of our estimates and those from IPCC 1994. For example, we estimate N<sub>2</sub>O emissions from agricultural N that flows from rivers into oceans. Part of the 'anthropogenic'  $N_2O$  is termed as 'natural' oceanic  $N_2O$  in the IPCC 1994 estimates. Also, because of the emissions of ammonia and nitrogen oxides from agricultural sources, and their eventual redeposition onto 'natural' systems,  $N_2O$  emissions from natural ecosystems may include  $N_2O$  derived from N evolved from agricultural soils and redeposited onto forest and grasslands. Exact quantification for these overlaps is difficult, and may have resulted in double counting of some sources. These new estimates suggest that the missing  $N_2O$  source discussed in earlier IPCC reports (Table 1) is likely to be a biogenic (agricultural) one.

#### Acknowledgements

We thank Bo Lim and collegues of OECD/IPCC the following participants of the workgroup who either provided information and suggestions and/or participated in the OECD/IPCC workshop for their contributions to this paper: L Bakken, P Bielek, S Bogdanov, Y Bonduki, A F Bouwman, R A Delmas, F J Dentener, R Francisco, J Freney, S Frolking, P Groffman, O Heinemeyer, R Karaban, L Klemedtsson, P A Leffelaar, E Lin, K Minami, W J Parton, DC Parashar, R Scholes, R Sherlock, K Smith, H g van Faassen, E Veldkamp, G L Velthof, G X Xing.

## References

- Benckiser G, Elts R, Linn A, Lorch HJ, Suemer E, Weiske A & Wenzhoefer F (1996) N<sub>2</sub>O emissions from different cropping systems and from aerated, nitrifying tanks of a municipal waste waters treatment plant. Biol Fertil Soils 23: 257–265
- Berounsky VM & Nixon SW (1993) Rates of nitrification along and estuarine gradient in Narrangasett Bay. Estuaries 16(4): 718–730
- Berounsky VM & Nixon SW (1985) Eutrophication and the rate of net nitrification in a coastal marine ecosystem. Estuarine Coastal and Shelf Science 20: 773–781
- BKH (1994) Studie naar de vorming van N<sub>2</sub>O in rioolwaterzuiveringsinstallaties. Literatuuronderzoek en orienterende metingen. BKH Report RO181018/2350I/2 Delft, the Netherlands
- Bouwman AF (1996) Direct emission of nitrous oxide from agricultural soils. Nutrient Cycling in Agroecosystems 46: 53–70
- Bouwman AF (1995) Compilation of a global inventory of emissions of nitrous oxide. Thesis Landbouwuniversiteti Wageningen. 143 p
- Bouwman AF (1994) Method to estimate direct nitrous oxide emisions from agricultural soils. Report 773004004. National Institute of Public Health and Environmental Protection, Bilthoven, the Netherlands 28 p
- Bouwman AF, Olivier JGJ & Van der Hoek KW (1995) Uncertainties in the global sourced distribution of nitrous qoxide. J Geophys Res 100: 2785–2800

- Bouwman AF & Van der Hoek KW (1991) Analysis of soil and water borne emissions of nitrous oxide and methane in the Netherlands. RIVM Report no. 736301010. National Institute of Public Health and Environmental Protection, Bilthoven, the Netherlands
- Bouwman AF & Sombroek WG (1990) Inputs to climatic change by soils and agriculture related activities: Present status and possible future trends. In: Scharpenseel HW, Schomaker M & Ayoub A (eds) Soils on a Warmer Earth. Developments in Soil Science, Vol 20 p 15–30. Elsevier, Amsterdam
- Bowden RD, Melillo JM & Steudler PA (1991) Effects of nitrogen additions on annual nitrous oxide fluxes from temperate forests soils in the northeastern United States. J Geophys Res 96: 9321–9328
- Bowden WB & Bormann FH (1986) Transport and loss of nitrous oxide in soil water afterforest clear-cutting. Science 233: 867–869
- Brumme R & Beese F (1992) Effects of liming and nitrogen fertilization on emission of  $CO_2$  and  $N_2O$  from a temperate forest. J Geophys Res 97: 12851–12858
- Burton CH, Sneath RW & Farrent JW (1993) Emissions of nitrogen oxide gases during aerobic treatment of animal slurries. Bioresource Technology 45: 233–235
- Carran RA, Theobald PW & Evans JP (1995) Emission of nitrous oxide from some grazed pasture soils in New Zealand. Aust J Soil Res 33: 341–352
- Christensen PB & Soerensen J (1988) Denitrification in sediment of lowland streams: regional and seasonal variation in Gelbaek and Rabis Beak, Denmark, FEMS Microbiol Eco 153: 335–344
- Christensen PB, Nielsen LP, Revsbech NP & Sorensen J (1989) Microzonation of denitrification activity in stream sediments as studies with a combined oxygen and nitrous oxide microsensor. Appl Environ Microbiol 55: 1234–1241
- Cole V, Cerri C, Minami K, Mosier A, Rosenberg N & Sauerbeck D et al. (1996) Chapter 23. Agricultural Options for Mitigation of Greenhouse Gas Emissions. pp 745–771. In: Watson RT, Zinyowera MC & Moss RH (eds) Climate Change 1995. Impacts, Adaptations and Mitigation of Climate Change: Scientific-Technical Analyses. Published for the Intergovernmental Panel on Climate Change. Cambridge University Press, Cambridge
- Cooper AB & Cooke JG (1984) Nitrate loss and transformation in 2 vegetated headwater streams. N Z J Mar Freshwater Res 18: 441–450
- Corrie MD, Van Kessel C & Pennock DJ (1996) Landscape and seasonal patterns of nitrous oxide emissions in a semiarid region. Soil Sci Soc Am J 60: 1806–1815
- Czepiel P, Crill P & Harriss R (1995) Nitrous oxide emissions from municipal wastewater treatment. Environ Sci Technol 29(9): 2352–2356
- Danso SKA (1995) Sustainable agriculture. The role of biological nitrogen fixing plants. In: IAEA (ed) Nuclear Techniques in Soil-Plant Studies for Sustainable Agriculture and Environmental Preservation pp 205–224. IAEA, Vienna
- Debruyn W, Lissens G, Van Rensbergen J & Wevers M (1994) Nitrous oxide emissions from waste water. In: Van Ham J et al. (eds) 1994: 159–165
- De Klein CAM & Van Logtestijn RSP (1994) Denitrification and N<sub>2</sub>O emission from urine-affected grassland soil. Plant and Soil 163: 235–242
- DeMore WB, Sander SP, Golden DM, Hampson RF, Kurylo MJ, Howard CJ, Ravishankara AR, Kolb CE & Molina MJ (1994) Chemical kinetics and photochemical data for use in stratospheric modeling. JPL-NASA, Evaluation No. 11

- Dentener FJ & Crutzen PJ (1994) A three-dimensional model of the global ammonia cycle. J Atmos Chem 19: 331–369
- Dowdell RJ, Burford JR & Crees R (1979) Losses of nitrous oxide dissolved in drainage water from agricultural land. Nature 278: 342–343
- Duff JH, Triska FJ & Oremland RS (1984) Denitrification associated with stream periphyton: Chamber estimates from undisrupted communities. J Environ Qual 13(4): 514–518
- Duxbury JM, Bouldin DR, Terry RE & Tate RL III (1982) Emissions of nitrous oxide from soils. Nature 298: 462–464
- Duxbury JM & Mosier AR (1993) Status and issues concerning agricultural emissions of greenhouse gases. In: Kaiser HM & Drennen TE (eds) Agricultural Dimensions of Global Climate Change, pp 229–258. St. Lucie Press. Delray Beach, FL
- ECETOC (1994) Ammonia emissions to air in Western Europe. Technical Report No 62. Brussels, 196 p
- Erich MS, Bererie A, & Duxbury JM (1984) Activities of denitrifying enzymes in freshly sampled soils. Soil Sci 38: 25–32
- Eswaran H, Van den Berg E & Reich P (1993) Organic carbon in soils of the world. Soil Sci Soc Am J 57: 192–194
- FAO (1990a) Fertilizer Yearbook. Volume 39. FAO statistics series No. 95. FAO, Rome
- FAO (1990b) Fertilizer Yearbook. Volume 43. FAO statistics series No. 94. FAO, Rome
- Firestone MK & Davidson EA (1989) Microbiological basis of NO and N<sub>2</sub>O production and consumption in soil. In: Andreae MO & Schimel DS (eds) Exchange of Trace Gases between Terrestrial Ecosystems and the Atmosphere, pp 7–21. John Wiley and Sons Ltd., Chichester, UK
- Flessa H, Doersch P, Beese F, Koenig H & Bouwman AF (1996) Influence of cattle excrements on nitrous oxide and methane fluxes in pasture land J Environ Qual 25: 1366–1370
- Flessa H, Doersch P & Beese F (1995) Seasonal variation of N<sub>2</sub>O and CH<sub>4</sub> fluxes in differently managed arable soils in southern Germany, J Geophy Res 100: 23,115–23, 124
- Galbally IE et al. (1992) Proceedings of IGBP Workshop No. 14. Canberra, Oct. 3-5, 1990
- Goreau TJ, Kaplan WA, Wofsy SC, McElroy MB, Valois FW & Watson SW (1980) Production of NO<sub>2</sub>-and N<sub>2</sub>O by nitrifying bacteria at reduced concentrations of oxygen. Appl Environ Microbiol 30: 526–532
- Groenestein CM, Oosthoek J & van Faassen HG (1993) Microbial processes in deep-litter systems for fattening pigs and emissions of ammonia, nitrous oxide and nitric oxide. In: Verstegen MWA et al. (eds) Nitrogen Flow in Pig Production and Environmental Consequences, p 307–312. Proc. First Int. Symp. on Nitrogen Flow in Pig Production and Environmental Consequences. Pudoc Scientific Publ., Wageningen
- Groffman PM, Rice CW & Tiedje JM (1993) Denitrification in a tallgrass prairie landscape. Ecology 74: 855–862
- Guthrie TF & Duxbury JM (1978) Nitrogen mineralization and denitrification in organic soils. Soil Sci Soc Am J 42: 908–912
- Hanaki K, Hong Z & Matsuo T (1992) Production of nitrous oxide gas during denitrification of wastewater. Wat Sci Tech 26(5–6): 1027–1036
- Hanson GC, Groffman PM & Gold AJ (1994) Denitrification in riparian wetlands receiving high and low groundwater nitrate inputs. J Environ Qual 23: 917–922
- Haynes RJ & Williams PH (1993) Nutrient cycling and soil fertility in grazed pasture ecosystem. Advances in Agronomy 49: 119– 199
- Helder W & De Vries RTP (1983) Estuarine nitrite maxima and nitrifying bacteria (ems-Dollard Estuary). Neth J Sea Res 17: 1–18

- Hemond HF & Duran AP (1989) Fluxes of N<sub>2</sub>O at the sedimentwater and water-atmosphere boundaries of a nitrogen-rich river. Water Res Res 25: 839–846
- Hill AR (1979) Denitrification in the nitrogen budget of a river ecosystem. Nature 281: 291–292
- Hill AR (1981) Nitrate-nitrogen flux and utilization in a stream ecosystem during los summer flows. Can Geogr 25(3): 225–239
- Hill AR (1983) Nitrate-nitrogen mass balances for two Ontario rivers. In: Fontaine TD & Bartell SM (eds) Dynamics of Lotic Ecosystems. Ann Arbor Science, Michigan
- Hong Z, Hanaki K & Matsuo T (1993) Greenhouse gas-N<sub>2</sub>O production during denitrification in wastewater treatment. Water Sci Technol 7: 203–207
- Horrigan SG & Springer AL (1990) Oceanic and estuarine ammonium oxidation: Effects of light. Limnol Oceanogr 4: 316–328
- IPCC (1990) Climate Change: The IPCC Scientific Assessment.
  IPCC Working Group I Houghton JT, Jenkins GJ, & Ephraums JJ and WMO/UNEP (eds) Cambridge University Press, Cambridge, UK 365 pp
- IPCC (1992) Houghton JT et al. (eds) Climate Change (1992) The Supplementary Report to the IPCC Scientific Assessment. Published for the IPCC, Cambridge University Press
- IPCC (1995a) (eds) Houghton JT et al. Climate Change (1994) Radiative Forcing of Climate Change and An Evaluation of the IPCC IS92 Emission Scenarios. Published for the IPCC, Cambridge University Press, Cambridge, UK 337 pp
- IPCC (Intergovernmental Panel on Climate Change/Organization for Economic Cooperation and Development) (1995b) IPCC Guidelines for National Greenhouse Gas Inventories. OECD/OCDE, Paris
- IPCC (1997) (Intergovernmental Panel on Climate Change/Organization for Economic Cooperation and Development). Guidelines for National Greenhouse Gas Inventories. OECD/OCDE, Paris
- Jarvis SC & Pain BF (1994) Greenhouse gas emissions from intensive livestock systems: their estimation and technologies for reduction. Climate Change 27: 27–38
- Jenkins MC & Kemp WM (1984) The coupling of nitrification and denitrification in two estiarine sediments. Limnol Oceanogr 29: 609–619
- Jensen H B, Jorgensen K S & Soerensen J (1984) Diurnal variation of nitrogen cycling in coastal marine sediments II. Nitrous oxide emissions. Mar Biol 83: 177–183
- Jensen MH, Andersen TK & Soerensen J (1988) Denitrification in coastal bay sediment: regional and seasonal variation in Aarhus Bight, Denmark. Mar Ecol Prog Ser 48: 155–162
- Jorgensen KS, Jensen HB & Sorensen J (1984) Nitrous oxide production from nitrification and denitrification in marine sediments at low oxygen concentrations. Can J Microbiol 30: 1073–1078
- Kaspar HF & Tiedje JM (1981) Dissimilatory reduction of nitrate and nitrite in the bovine rumen: nitrous oxide production and effect of acetylene. Appl Environ Microbiol 41: 705–709
- Kaushik NK & Robinson JB (1976) Preliminary observations on nitrogen transport during summer in a small spring-fed Ontario stream. Hydrobiol 49(1): 59–63
- Kemp WM, Sampou P, Caffrey J & Mayer M (1990) Ammonium recycling versus denitrification in Chesapeake Bay sediments. Limnol Oceanogr 35: 1545–1563
- Koops JG, Oenema O & van Beusichem ML (1996) Denitrification in the top and subsoil of grassland on peat soils. Plant and Soil 184: 1–11
- Koops JG, Oenema O & van Beusichem ML (1997) Nitrogen loss from grassland on peat soils through nitrous oxide production. Plant and Soil 188: 119–130

- Kroeze C & Seitzinger SP (1998) Nitrogen inputs to rivers, estuaries and continental shelves and related nitrous oxide emissions in 1990 and 2050: A global model. Nutrient Cycling in Agroecosystems 52: 195–212
- Kroeze C (1994) Nitrous Oxide; Emission inventory and options for control in the Netherlands. Report Number 773001004. National Institute of Public Health and Environmental Protection, Bilthoven, the Netherlands. 163 p
- Kumarasinghe KS & Eskew DL (1991) Applications of N-15 isotope techniques in Azolla-rice studies. In: IAEA (ed) Stable Isotopes in Plant Nutrition, Soil Fertility and Environmental Studies, pp 169–178. IAEA, Vienna
- Li C, Frolking S & Frolking TA (1992) A model of nitrous oxide evolution from soil driven by rainfall events: 1. Model structure and sensitivity. J Geophys Res 97: 9759–9766
- Lipschultz F, Wofsy SC & Fox LE (1986) Nitrogen metabolism of the eutropic Delaware River ecosystem. Limnol Oceanogr 29: 84–98
- Martikainen PJ, Nykanen H, Crill P & Silvola J (1996) Effect of a lowered water table on nitrous oxide fluxes from northern peatlands. Nature 366: 51–53
- Minami K & Fukushi S (1984) Methods for measuring N<sub>2</sub>O flux from water surface and N<sub>2</sub>O dissolved in water from agricultural land. Soil Sci Plant Nutr 30: 495–502
- Minami K & Ohsawa A (1990) Emission of nitrous oxide dissolved in drainage water from agricultural land. In: Bouwman AF (ed) Soils and the Greenhouse Effect, pp 503–509. John Wiley & Sons New York
- Monaghan RM & Barraclough B (1993) Nitrous oxide and dinitrogen emissions from urine-affected soil under controlled conditions. Plant and Soil 151: 127–138
- Mosier AR (1994) Nitrous oxide emissions from agricultural soils. Fert Res 37: 191–200
- Mosier AR & Parton WJ (1985) Denitrification in a shortgrass prairie: A modeling approach. In: Caldwell DE, Brierley JA & Brierley CL (eds) Planetary Ecology, pp 441–452. Van Nostrand Reinhold Co., NY.
- Mosier AR & Bouwman AF (1993) Working group report: Nitrous oxide emissions from agricultural soils. In: van Amstel AR (ed) Methane and Nitrous Oxide: Methods in National Emission Inventories and Options for Control Proceedings. 00 343–346. National Institute of Public Health and Environmental Protection, Bilthoven, the Netherlands
- Mosier AR, Schimel DS, Valentine D, Bronson KF & Parton WJ (1991) Methane and nitrous oxide fluxes in native, fertilized, and cultivated grasslands. Nature 350: 330–332
- Mosier AR, Duxbury JM, Freney JR, Heinemeyer O & Minami K (1996) Nitrous oxide emissions from agricultural fields: Assessment, measurement and mitigation. Plant and Soil 181: 95–108
- Mosier AR, Duxbury JM, Freney JR, Heinemeyer O & Minami K (1997) Mitigating agricultural emissions of nitrous oxide. Climatic Change (in press)
- Muller C, Sherlock RR & Williams PH (1995a) In-field method to determine N<sub>2</sub>O and N<sub>2</sub> emissions from denitrification and N<sub>2</sub>O emissions from nitrification from urine-affected intensive pasture soil. Submitted to Soil Biology and Biochemistry
- Muller C, Sherlock RR & Williams PH (1995b) Factors influencing N<sub>2</sub>O emission from pasture soil affected by animal urine or ploughing. Submitted to European Journal of Soil Science
- Nevison CD, Esser G & Holland EA (1996) A Global Model of Changing N<sub>2</sub>O Emissions from Natural and Perturbed Soils. Climatic Change 32: 327–378

- Nobre AD (1994) Nitrous oxide emissions from tropical soils. Thesis. Univ. New Hampshire, Durham, NH. 141 pp
- Nykanen, Alm HJ, Loeng K, Silvola J & Martikainen PJ (1995) Emissions of CH<sub>4</sub>, N<sub>2</sub>O and CO2 from virgin fen and fen drained for grassland in Finland. J Biogeography 22: 351–357
- OECD/OCDE (1991) Estimation of Greenhouse Gas Emissions and Sinks. Final report from the OECD Experts Meeting, 18-21 Feb., 1991. Prepared for Intergovernmental Panel on Climate Change. Revised August, 1991
- Oenema O, Velthof GL, Yamulki S & Jarvis SC (1997) Nitrous oxide emission from grazed grassland. Soil Use Manage 13: 1–9
- Oenema O & Velthof GL (1993) Denitrification in nitric-acidtreated cattle slurry during storage. Netherlands J Agric Sci 41: 63–80
- O'Hara GW & Daniel RM (1985) Rhizobial denitrification: a review. Soil Biol Biochem 17: 1-9
- Oonk H & Kroeze C (1998) Nitrous oxide (N<sub>2</sub>O) emissions and control. In: RA Meyers (ed) Encyclopedia of Environmental Analysis and Remediation. John Wiley & Sons Inc (In Press).
- Owens NJP (1986) Estuarine nitrification: a naturally occurring fluidized bed reaction? Estuarine Coastal and Shelf Science 33: 31–44
- Parashar DC (1991) Measurement of greenhouse gas emissions in India. In: Impact of Global Climate Change on Photosynthesis and Plant Productivity, pp 625–640. Oxford & IBH Publishing Co., New Delhi
- Parton WJ, Mosier AR, Ojima DS, Valentine DW, Schimel DS, Weier K & Kulmala AE (1996) Generalized model for  $N_2$  and  $N_2$ O production from nitrification and denitrification. Global Biogeochem Cycles 10: 401–412
- Peoples MB, Herridge DF & Ladha JK (1995) Biological nitrogen fixation: An efficient source of nitrogen for sustainable agricultural production? Plant and Soil 174: 3–28
- Poggemann S, Weissbach F & Kuenzel U (1995) N<sub>2</sub>O-Freisetzung von Exkrementflecken auf dem Weideland. Abstract. VDLUFA, Garmisch-Partenkirchen. p 275
- Pollaris L (1994) Emission of nitrous oxide from horticultural substrates. (In Dutch) M. Sc. Thesis. Faculty of Agricultural and Applied Biological Sciences. University of Gent. 102 p
- Postma R, Oenema O, Bussink DW, Heinen M, & Van Mollenbroek J (1994) Gaseous Nitrogen Emissions from Lettuce Grown in Irrigated Sand Beds in Glasshouse Horticulture, pp 28–34. Meststoffen
- Potter CS, Matson PA, Vitousek PM & Davidson EA (1996) Process modeling of controls on nitrogen trace gas emissions from soils worldwide. J Geophys Res 101: 1361–1377
- Robertson GP (1993) Fluxes of nitrous oxide and other nitrogen trace gases from intensively managed landscapes. In: Harper LA, Mosier AR, Duxbury JM & Rolston DE (eds) Agroecosystem Effects on Radiatively Important Trace Gases and Global Climate Change, pp 95–108. ASA Special Publication No. 55. Amer Soc Agron, Madison, WI
- Robinson JB, Whiteley HR, Stammers W, Kaushik NK & Sain P (1979) The fate of nitrate in small streams and its management implications, pp 247–259. In: Proc 10th Annu Agric Waste Manage Conf Cornell Univ
- Ronen D, Margaritz M & Alman E (1988) Contaminated aquifers are a forgotten component of the global N<sub>2</sub>O budget. Nature 335: 57–59
- Ryden JC (1981)  $N_2O$  exchange between a grassland soil and the atmosphere. Nature 292: 235–365
- Ryden JC (1983) Denitrification loss from a grassland soil in the field receiving different rates of nitrogen as ammonium nitrate. J Soil Sci 1983: 355–365

- Safley LM, Casada ME, Woodbury JW & Roos KW (1992) Global methane emissions from livestock and poultry manure. USEPA Report No. 400/1-91/048. Office of Air and Radiation, Washington, D.C. 68 p with 145 p annexes
- Schepers JS & Mosier AR (1991) Accounting for nitrogen in non-equilibrium soil-crop systems. In: Follett RF, Keeney DR & Cruse RM (eds) Managing Nitrogen for Groundwater Quality and Farm Profitability, pp 125–138. Soil Science Society of America, Inc. Madison, Wisconsin
- Schimel DS, Parton WJ, Adamsen FJ, Woodmansee RG, Senft RL & Stillwell MA (1986) The Role of Cattle in the Volatile Loss of Nitrogen from a Shortgrass Steppe. Biogeochemistry 2: 39–52
- Seitzinger SP (1988) Denitrification in freshwater and coastal marine ecosystems: ecological and geochemical implications. Limnol Oceanogr 33: 702–724
- Seitzinger SP (1990) Denitrification in aquatic sediments. In: Revsbech NP & Sorensen J (eds) Denitrification in Soil and Sediment, pp 301–312. Plenum Press, New York
- Seitzinger SP & Kroeze C Estimates of the spatial distribution of N inputs, nitrous oxide production and denitrification in freshwater and coastal marine ecosystems. (in preparation)
- Sherlock RR & Goh KM (1983) Initial emission of nitrous oxide from sheep urine applied to pasture soil. Soil Biol Biochem 15: 615–617
- Sibbesen E & Lind AM (1993) Loss of nitrous oxide from animal manure dungheaps. Acta Agriculture Scandianavia Section B: Soil Plant Sci 43: 16–20
- Smith CJ, DeLaune RD & Patrick WH Jr (1985) Fate of riverine nitrate entering an estuary: I. Denitrification and nitrogen burial. Estuaries 8: 15–21
- Spoelstra SF (1985) Nitrate in silage. Grass Forage Sci 40: 1–11
   Steffens G & Vetter H (1990) Neue Faustzahlen ueber Nachrstoffgehalte und Naehrstoffanfall. Landwirtsschaftsblatt Weser-Ems.
   Vol. 3
- Swank WT & Caskey WH (1982) Nitrate depletion in a secondorder mountain stream. J Environ Qual 11: 581–584
- Thornton FC & Valente RJ (1996) Soil emissions of nitric oxide and nitrous oxide from no-till corn. Soil Sci Soc Am J 60: 1127–1133
- Tiedje JM (1988) Ecology of denitrification and dissimilatory nitrate reduction to ammonia. In: Zehnder AJB (ed) Biology of Anaerobic Microorganisms, pp 179–243. Wiley, New York
- Ueda S, Ogura N & Wada E (1991) Nitrogen stable isotope ratio of groundwater  $N_2O$ . Geophys Res Lett 18 (8): 1449–1452
- Ueda S, Ogura N & Yoshinari T (1993) Accumulation of nitrous oxide in aerobic groundwaters. Water Res 27 (12): 1787–1792
- Van Aardenne JA (1996) Uncertainty and sensitivity analysis of an IPCC/OECD model for estimating N<sub>2</sub>O emissions from agricultural soils. AV milieusysteemanalyse H250-705, 29 November, 1996. Center for Environment and Climate Change Research, Wageningen Agricultural University, Wageningen, the Netherlands, 32 p

- Van Bochove E, Jones HG, Pelletier P & Prevost D (1996) Emission of N<sub>2</sub>O from agricultural soil under snow cover: A significant part of N budget. Hydrological Processes 10: 1545–1549
- Van Cleemput O, Vermoesen A, De Groot CJ & Van Ryckeghem K (1994) Nitrous oxide emission out of grassland. Environ Monitor Assessm 31: 145–152
- Van der Hoek KW (1994) Berekeningsmethodiek ammoniamemissie in Nederland voor de jaren 1990, 1991 en 1992. RIVM report No. 773004003
- Van Kessel JP (1977) Removal of nitrate from effluent following discharge on surface water. Water Res 11: 533–537
- Veldkamp E & Keller M (1997) Nitrogen oxide emissions from a banana plantation in the humid tropics. J Geophys Res (in press)
- Velthof GL, Brader AB & Oenema O (1996a) Seasonal variations in nitrous oxide losses from managed grasslands in the Netherlands. Plant and Soil 181: 263–274
- Velthof GL, Jarvis SC, Stein A, Allen AG & Oenema O (1996b) Spatial variability of nitrous oxide fluxes in mown and grazed grasslands on a poorly drained clay soil. Soil Biol Biochem 28: 1215–1225
- Velthof GL & Oenema O (1993) Orienterende lachgasemissiemetingen in RWZI's. NMI-Report 93.263. Wageningen. 6 pp
- Velthof GL & Oenema O (1994) Effect of nitrogen fertilizer type and urine on nitrous oxide flux from grassland in early spring. In: Mannetje L & Frame J (eds) Grassland and Society, pp 458–462. Proceedings of the 15th General Meeting of the European Grassland Federation
- Velthof GL & Oenema O (1995) Nitrous oxide fluxes from grassland in the Netherlands: II. Effects of soil type, nitrogen fertilizer application and grazing. European J Soil Sci 46: 541–549
- Vetter H, Klasink A & Steffens G (1989) Mist und Guelledue Ingung nach Mass. VDLUFA-Schriftenreihe. 19: 41–66
- Wagner-Riddle C & Thurtell GW (1998) Nitrous oxide emissions from agricultural fields during winter and spring thaw as affected by management practices. Nutrient Cycling in Agroecosystems 52: 151–163
- Whitehead DC (1970) The role of nitrogen in grassland productivity. Bulletin 48 of Commonwealth Agricultural Bureau, Hurley, UK
- WHO/UNEP (1989) Global Pollution and Health. Results of health related environmental monitoring, World Health Organization. Geneva and United Nations Environment Programme, Nairobi, Yale Press, London

Appendix 1. Default Values for Percentage of Manure N Produced in Different Animal Waste Management Systems in Different World Regions (from Safley et. al., 1992)

Region	Type of Animal	Percent of Manure Production per Animal Waste Management Systems							
		Anaerobic Lagoon	Liquid System	Daily Spread	Solid Storage and Drylot	Pasture Range and Paddock	Used Fuel	Other System	
North America	Non-dairy Cattle (D)	0	1	0	14	84	0	1	
	Dairy Cattle	10	23	37	23	0	0	7	
	Poultry (E)	5	4	0	0	1	0	90	
	Sheep	0	0	0	2	88	0	10	
	Swine	25	50	0	18	0	0	6	
	Other animals (F)	0	0	0	0	92	0	8	
Western Europe	Non-dairy Cattle (D)	0	55	0	2	33	0	9	
_	Dairy Cattle	0	46	24	21	8	0	1	
	Poultry (E)	0	13	0	1	2	0	84	
	Sheep	0	0	0	2	87	0	11	
	Swine	0	77	0	23	0	0	0	
	Other animals (F)	0	0	0	0	96	0	4	
Eastern Europe	Non-dairy Cattle (D)	8	39	0	52	0	0	1	
	Dairy Cattle	0	18	1	67	13	0	0	
	Poultry (E)	0	28	0	0	1	0	71	
	Sheep	0	0	0	0	73	0	27	
	Swine	0	29	0	0	27	0	45	
	Other animals (F)	0	0	0	0	92	0	8	
Oceania	Non-dairy Cattle (D)	0	0	0	0	100	0	0	
o commu	Dairy Cattle	0	0	0	0	100	0	0	
	Poultry (E)	0	0	0	0	3	0	98	
	Sheep	0	0	0	0	100	0	0	
	Swine	55	0	0	17	0	0	28	
	Other animals (F)	0	0	0	0	100	0	0	
Latin America	Non-dairy Cattle (D)	0	0	0	0	99	0	1	
Laun America	Dairy Cattle	0	1	62	1	36	0	0	
	Poultry (E)	0	9	0	0	42	0	49	
	Sheep	0	0	0	0	100	0	0	
	Swine	0	8	2	51	0	0	40	
	Other animals (F)	0	0	0	0	99	0	1	
Africa	Non-dairy Cattle (D)	0	0	1	3	96	0	0	
Affica	Dairy Cattle	0	0	12	0	83	0	5	
	Poultry (E)	0	0	0	0	81	0	3 19	
	•	0	0	0	1	99	0	19	
	Sheep Swine	0	7	0	93	0	0	0	
	Other animals (F)	1	0	0	0	99	0		
Noon Foot and	Other animals (F)	1	U	U	U	99	U	1	
Near East and	Non doing Cattle (D)	0	0	2	0	77	10	2	
Mediterranean	Non-dairy Cattle (D)	0	0	2	0	77	18	2	
	Dairy Cattle	0	0	3	3	77 71	18	0	
	Poultry (E)	0	1	0	0	71	0	28	
	Sheep	0	0	0		100	0	0	
	Swine	0	32	0	68	0	0	0	
	Other animals (F)	0	0	0	0	100	0	0	

248 Appendix 1. Continued

Region	Type of Animal	Percent of Manure Production per Animal Waste Management Systems							
		Anaerobic Lagoon	Liquid System	Daily Spread	Solid Storage and Drylot	Pasture Range and Paddock	Used Fuel	Other System	
Asia and Far East	Non-dairy Cattle (D)	0	0	16	14	29	40	0	
	Dairy Cattle	6	4	21	0	24	46	0	
	Poultry (E)	1	2	0	0	44	1	52	
	Sheep	0	0	0	0	83	0	17	
	Swine	1	38	1	53	0	7	0	
	Other animals (F)	0	0	0	0	95	0	5	

<sup>(</sup>D) Includes buffalo.

<sup>(</sup>E) Includes chickens, turkeys and ducks.(F) Includes goats, horses, mules, donkeys and camels.